

1 **Appendix G Biological Resources –**

2 **Fisheries**

3 This appendix describes the environmental setting for fisheries resources.

4 **G.1 Environmental Setting**

5 **G.1.1 Regional Setting**

6 ***Historical Habitat***

7 Typical of Central Valley rivers and a semiarid climate, the natural or “unimpaired” flow
8 regime of the San Joaquin River historically varied greatly in the magnitude, timing,
9 duration, and frequency of streamflows, both interannually and seasonally (San Joaquin
10 River Restoration Program [SJRRP] 2011a). Streamflow variability created conditions
11 that partially helped create and sustain multiple salmonid life history strategies and life
12 history phases of numerous other resident and anadromous native fish and other aquatic
13 species.

14 The San Joaquin River originates in the Sierra Nevada at an elevation greater than
15 13,000 feet above mean sea level (SJRRP 2011a). It rapidly descends and exits
16 mountainous terrain in the area now occupied by Friant Dam. The San Joaquin River
17 downstream from Friant Dam consists of a deeply incised channel that discharges to the
18 valley floor near Gravelly Ford. Before the influx of settlers in the 1860s, and subsequent
19 agricultural development, the San Joaquin River and its main tributaries meandered
20 across alluvial fans, deposits of river sediments resulting from a decrease in velocity,
21 along the main axis of the San Joaquin Valley floor in their natural state. Historically, the
22 San Joaquin River in Reach 1 was moderately sinuous with a gravel bed and planform
23 morphology with numerous split channels, side channels, and high flow scour channels
24 (McBain and Trush 2002). In Reach 2 the river transitioned into being sand bedded with
25 a meandering morphology with large sinuosity and a single primary channel (McBain and
26 Trush 2002). High flow scour channels at the downstream end of Reach 2 transported
27 high flows south to Fresno Slough which were then conveyed back to the San Joaquin
28 River at Mendota (McBain and Trush 2002). Reach 3 had similar morphology to Reach 2
29 with large exposed point bars and riparian vegetation present on the top of the point bars
30 and on the floodplains (McBain and Trush 2002).

31 Near Mendota, the San Joaquin River merged with Fresno Slough, a wider and deeper
32 waterway than the San Joaquin River (SJRRP 2011a). Fresno Slough was part of an
33 intricate slough system that exchanged water between the Tulare Lake Basin and the San
34 Joaquin River. Downstream from Mendota, in the present area of the Reach 4B/ESB
35 Project study area, the San Joaquin River was a meandering sand-bedded channel with
36 numerous anabranching sloughs with base flows being conveyed by both the San Joaquin

1 River channel and the sloughs (McBain and Trush 2002). Narrow riparian levees
2 provided moderate confinement of the river on both banks, with large areas of tule marsh
3 flood basins being present past the riparian levees (The Bay Institute 1998, McBain and
4 Trush 2002). Oxbow lakes and off-channel ponds within the flood basins were likely
5 present (McBain and Trush 2002). The flood basins extended for miles on both sides of
6 the San Joaquin River in Reach 4B (McBain and Trush 2002). Channel migration and
7 avulsion were likely very slow and infrequent due to the low sediment supply, as a result
8 of deposition in upstream reaches, and low stream energy as high flows spilled over the
9 narrow riparian levees into the flood basins (McBain and Trush 2002). With the limited
10 channel confinement provided by the riparian levees, overbank inundation of the flood
11 basins probably occurred most years and was of long duration, on the order of months
12 (McBain and Trush 2002). The prolonged inundation of sloughs and flood basins likely
13 provided high flow refugia and rearing habitat for juvenile salmonids and other native
14 fishes (McBain and Trush 2002).

15 Although historic water quality data (i.e., data from before construction of Friant Dam)
16 are not available, the river provided sufficient water quality conditions for native fish,
17 including anadromous salmonids (SJRRP 2011a). Cold, clear snowmelt runoff flowing
18 from the granitic upper basins of the southern Sierra Nevada provided optimal conditions
19 for freshwater life history stages of salmonids in the upper San Joaquin River and its
20 tributaries and for invertebrate production, the primary food resource for salmonids. The
21 abundant cold water in the upper San Joaquin River basin had high (saturated)
22 concentrations of dissolved oxygen (DO), low salinity, and neutral pH levels. Levels of
23 suspended sediment and turbidity likely were relatively low, even during high runoff
24 events, because of the upper basin's mainly granitic geology and relatively low rates of
25 primary productivity (algae growth). In the Reach 4B/ESB Project area, primary
26 productivity likely increased historically as a result of the river meandering through
27 sloughs, flood basins, and long, slow pools with minimal shading from riparian
28 vegetation. However, the extensive tule marshes in the flood basins may have provided
29 extensive shade in locations where the tules were dense.

30 The San Joaquin River in Reach 4B was historically a gaining reach with shallow
31 groundwater being very close or above the river surface and discharging to the river and
32 surrounding marshes (McBain and Trush 2002). During periods of low surface flow, the
33 shallow unconfined aquifer of the San Joaquin Valley trough would contribute significant
34 baseflows to the San Joaquin River in Reach 4B (McBain and Trush 2002). The shallow
35 groundwater pumping close to the river has changed portions of the San Joaquin River
36 within Reach 4B from a gaining reach to a losing reach (McBain and Trush 2002).

37 ***Existing Habitat***

38 The San Joaquin Valley, part of the San Joaquin Basin, and the associated Tulare Basin,
39 once had a wide variety of terrestrial and aquatic habitats that provided rich resources for
40 Native Americans and early settlers (Brown 2000). However, as the San Joaquin Valley
41 was converted to agricultural land use, native ecological communities declined. On the
42 valley floor, invasive species, intensive agricultural activity, and increasing urbanization,
43 have resulted in changes to water quality and aquatic habitats. Invasive species have
44 caused changes in aquatic and riparian plant communities resulting in reduced habitat

1 quality. Intensive pesticide and fertilizer use, which enter surface waters in various ways,
2 has impacted water quality (Kuivila and Foe 1995, Domagalski et al. 1997, Kratzer and
3 Shelton 1998, Brown et al. 1999). Pesticide concentrations sometimes reach
4 concentrations acutely toxic to sensitive invertebrates (Kuivila and Foe 1995).
5 Agricultural return flows also may contain high concentrations of dissolved solids
6 (salinity) and trace elements (Saiki 1984, see Brown 2000) that can degrade water
7 quality. Clearing of land for agriculture or flood control activities has resulted in the loss
8 of over 90% of wetland and riparian habitat (Brown 2000).

9 Today, water resource systems of the San Joaquin River region are among the most
10 constrained in the nation as managers try to meet water supply, water quality, flood
11 control, ecosystem, and recreation objectives (Brekke et al. 2004). During the irrigation
12 season (usually March to October), irrigation water from the Delta-Mendota Canal and
13 return flows from irrigated fields usually contribute most of the discharge in downstream
14 portions of the San Joaquin River above the Merced River confluence (Reaches 3-5)
15 (Saiki and Palawski 1990). In contrast, the west-side tributaries, some of which originate
16 on the San Joaquin Valley floor, derive most of their discharge from groundwater
17 seepage and irrigation return flows. The section of San Joaquin River between Friant
18 Dam and the Merced River confluence (i.e., Reaches 1A through 5) provides generally
19 poor fish habitat conditions (SJRRP 2010a). Physical barriers and reaches with poor
20 water quality or no surface flow have reduced habitat connectivity. Under current
21 operations, approximately 60 miles of the San Joaquin River are dewatered for the
22 majority of the year. Tributaries to these reaches support little or no available spawning
23 habitat for anadromous salmonids and lamprey and, under certain conditions, potentially
24 create straying opportunities that hinder the ability of fish to complete their life cycles,
25 especially adult salmonids. Habitat complexity between Friant Dam and the confluence
26 with the Merced River is reduced, with limited secondary habitat (e.g., side channels and
27 floodplains) or instream habitat structure, and contains highly altered riparian vegetation.
28 Bypasses in these reaches receive water sporadically, as necessary for flood control.

29 **G.1.2 Reach 4B/ESB Project Area Setting**

30 The Reach 4B/ESB Project area includes Reaches 4B1 and 4B2, a 32.5-mile stretch of
31 the San Joaquin River, the Middle Eastside Bypass, the Lower Eastside Bypass, and the
32 Mariposa Bypass in Merced County, California (see Figure 1-2). A wide variety of
33 aquatic and upland habitats occur within the Reach 4B/ESB Project area, but the habitats
34 that most directly impact fishery resources include: riverine/open water, lacustrine,
35 freshwater emergent wetland, seasonal wetland (if connected hydrologically to the main
36 channel so that fish have access), riparian/willow scrub, and valley foothill riparian. The
37 distribution, species compositions, and abundance of these habitats are described in detail
38 in Chapter 6 of this EIS (Biological Resources-Vegetation and Wildlife).

39 **Reach 4B**

40 Reach 4B of the San Joaquin River begins at the Sand Slough Control Structure (River
41 Mile [RM] 168.5) and extends downstream to the confluence of the Eastside Bypass and
42 San Joaquin River (RM 136) (see Figure 1-2). Reach 4B has been further divided into
43 two sub-reaches, Reach 4B1 and Reach 4B2. Reach 4B1 begins at the Sand Slough

1 Control Structure and continues to the Mariposa Bypass, and Reach 4B2 extends from the
2 Mariposa Bypass to the confluence of the Eastside Bypass and the San Joaquin River.

3 The section of river directly upstream of Reach 4B (Reach 4A) is dry in most months
4 because all flows in the San Joaquin River are diverted at Sack Dam to the Arroyo Canal.
5 Any flows reaching the Sand Slough Control Structure are diverted to the Eastside
6 Bypass via the Sand Slough Control Structure, leaving Reach 4B1 dry, with the exception
7 of agricultural return flows, local runoff, natural pooling due to shallow ground water in
8 wet years, and when it is used to convey water by land owners (SJRRP 2010a). As a
9 result, the Reach 4B1 channel has become poorly defined and has filled in with dense
10 vegetation and other fill material. In addition, Reach 4B1 is confined by
11 anthropogenically modified narrow levees. Reach 4B2 begins at the confluence of the
12 Mariposa Bypass, where flood flows in the bypass system rejoin the main stem of the San
13 Joaquin River, and this reach extends to the confluence of the Eastside Bypass (SJRRP
14 2011b). As a result of differences in manmade levee configuration, Reach 4B2 contains
15 wider floodplains and a more sinuous channel, including side channels and oxbows,
16 because of a wider levee configuration than Reach 4B1. Additionally, it contains vast
17 areas of grasslands and riparian vegetation stands. A portion of Reach 4B2 flows through
18 the San Luis National Wildlife Refuge (NWR), which is managed to support a wide
19 variety of native plant and animal species. Unlike Reach 4B1, Reach 4B2 is perennially
20 wet because of agricultural return flow (SJRRP 2010a).

21 ***Eastside Bypass and Mariposa Bypass***

22 The study area for the Reach 4B/ESB Project also includes the Eastside and Mariposa
23 bypasses. The Eastside Bypass extends from the confluence of Ash Slough and
24 Chowchilla Bypass to the confluence with the San Joaquin River at the head of Reach 5.
25 In the Grasslands Wildlife Management Area, riparian trees and shrubs have a patchy
26 distribution along the banks of the Eastside Bypass. The Lower Eastside Bypass has some
27 side channels and sloughs that support remnant patches of riparian vegetation. Outside of
28 the refuge areas, the Eastside Bypass is managed for flood conveyance and does not
29 currently support extensive riparian habitat. The Mariposa Bypass conveys flows from
30 the end of the Middle Eastside Bypass to the San Joaquin River at the upstream end of
31 Reach 4B2. The Mariposa Bypass is also managed for flood conveyance and does not
32 currently support riparian habitat. The bypasses are routinely cleared of vegetation to
33 maintain flood capacity and are regularly used to dispose of agricultural drain water.

34 The flood season for the Lower San Joaquin Levee District (LSJLD) typically lasts from
35 November 15 to June 15 of each water year, with rainfall contributing to higher flows
36 during the early part of the flood season, and snowmelt contributing to flows at the later
37 part of the flood season.

38 Key flood control structures within the study area include the Reach 4B Headgate on the
39 San Joaquin River at the beginning of Reach 4B1, the Sand Slough Control Structure at
40 the beginning of the Middle Eastside Bypass, the Eastside and Mariposa bypass control
41 structures where the Middle Eastside Bypass transitions to the Lower Eastside Bypass,
42 and the Mariposa Drop Structure at the end of the Mariposa Bypass near the confluence
43 with the San Joaquin River at the upstream end of Reach 4B2 (SJRRP 2011b).

1 **G.1.3 Environmental Stressors**

2 This section describes the major environmental stressors currently affecting native fish
3 species in the San Joaquin River. Stressors are defined as physical, chemical, or
4 biological perturbations to a system that adversely affect ecosystem processes, habitats,
5 and species (SJRRP 2010b). The following summarizes information from a literature
6 review of overall San Joaquin River Restoration Area (Restoration Area) stressors
7 (SJRRP 2010b, SJRRP 2011a), and findings from recent SJRRP investigations. When
8 SJRRP actions enable anadromous salmonids to use the Reach 4B/ESB Project area then
9 many of these stressors will also occur within the project area or affect fish in upstream
10 or downstream locations after they have passed through Reach 4B/ESB.

11 **Disease**

12 The fish diseases in downstream locations may occur in Reach 4B/ESB once fish passage
13 improvement allows Chinook Salmon and other fishes to consistently use and migrate
14 through the reach. The United States Fish and Wildlife Service (USFWS) conducted a
15 survey of the health and physiological condition of juvenile fall-run Chinook salmon
16 (*Oncorhynchus tshawytscha*) in the San Joaquin River and its primary tributaries, the
17 Stanislaus, Tuolumne, and Merced rivers, during spring 2000 and 2001 (Nichols and
18 Foott 2002). *Renibacterium salmoninarum*, the causative agent of bacterial kidney
19 disease (BKD), was detected in naturally produced juveniles caught in rotary screw traps
20 from the Stanislaus and Tuolumne rivers and juveniles caught with a Kodiak trawl at
21 Mossdale in the San Joaquin River. No gross clinical signs of BKD were seen in any of
22 the fish examined. However, these low-level infections might remain active after infected
23 fish enter the ocean where clinical symptoms might develop.

24 Proliferative kidney disease (PKD) was detected in both natural and hatchery juveniles
25 from the Merced and main stem San Joaquin rivers in 2000 and 2001 (Nichols and Foott
26 2002), and in natural juveniles from the Merced River in 2002 (Nichols 2002). The
27 myxozoan parasite *Tetracapsula bryosalmonae*, which causes PKD, was detected in the
28 kidney samples of only 2 percent of juvenile Merced River fish sampled in April 2000,
29 but 90 percent of April 2001, 100 percent of May 2001, and 51 percent of April 2002
30 samples. Heavy infections were observed in 22 percent of samples in 2002 (Nichols
31 2002). These data suggest that the incidence of pathogen infection is low in above-normal
32 water years, such as 2000, compared to dry water years such as 2001 and 2002. PKD has
33 been described at the Merced River Fish Hatchery since the 1980s and in California since
34 at least 1966. It compromises fish swimming, saltwater entry performance, and disease
35 resistance (Nichols and Foott 2002). Nichols and Foott (2002) suggest that PKD could be
36 a significant contributor to mortality in natural fish.

37 The pathogen *Ceratonova* (previously *Ceratomyxa*) is present in the Central Valley, and
38 studies indicate it can cause high mortality rates in Chinook smolts (Hendrickson et al.
39 1989, Foott and Imrie 2016). This disease relies on tubifex worms for an intermediate
40 host, and the worms flourish in organic sediments. It is likely the worms multiply, and the
41 disease spreads in years when organic sediments are not flushed by high flows. There are
42 indications that mortality of smolts due to this disease increases in drought years and
43 decreases in wet years (Foott and Imrie 2017).

1 Whirling disease is found in salmonid populations in the Central Valley (NMFS 2014).
2 The disease is caused by the parasite *Myxobolus cerebralis*, which has a two-aquatic host
3 life cycle consisting of an oligochaete worm *Tubifex tubifex* and a salmonid fish
4 (Steinbach Elwell et al. 2009). Very young fish are the most vulnerable to whirling
5 disease with susceptibility decreasing with age and growth (Steinbach Elwell et al. 2009).
6 The response of salmonids to infection by *M. cerebralis* varies among genera, species,
7 strains, and individuals (Steinbach Elwell et al. 2009). Within the genus *Oncorhynchus*,
8 most species experience high prevalence and severity of disease, and high mortality rates
9 if exposed to a sufficient parasite dose when susceptible (Steinbach Elwell et al. 2009).
10 Highly susceptible species include rainbow trout with Chinook salmon being more
11 resistant (Steinbach Elwell et al. 2009). The clinical signs of whirling disease include:
12 1) whirling behavior resulting from spinal cord constriction and brain stem compression,
13 2) blackened tail caused by pressure on nerves that control pigmentation, 3) skeletal
14 deformities caused by cartilage damage and interference with normal bone growth, and
15 4) mortality as a result of direct physical damage or inability to feed or avoid predation
16 (Steinbach Elwell et al. 2009). In addition, infection can reduce fitness by decreasing
17 growth rate and reducing swimming performance (Steinbach Elwell et al. 2009).

18 **Habitat Degradation**

19 The San Joaquin River within the Restoration Area has a sediment budget imbalance as a
20 result of the elimination by Friant Dam of most sediment supply from the upper
21 watershed in combination with the modified flow regime and land use downstream from
22 Friant Dam (SJRRP 2010b). Loss of alluvial features in the Restoration Area has
23 contributed to the reduction in frequency of floodplain inundation, which has probably
24 caused a substantial reduction in potential food resources and predator refuge for juvenile
25 salmonids in the Restoration Area (SJRRP 2010b). The loss of flow and encroachment of
26 levees, structures, flood control, and farming practices have also contributed to the
27 reduction in floodplain presence and frequency of floodplain inundation. Historically,
28 these inundation areas (flood basins, shallow sloughs, and side channels) may have
29 provided excellent rearing opportunities for juvenile salmonids and other species
30 (Sommer et al. 2001, Sommer et al. 2005, Jeffres et al. 2008, Limm and Marchetti 2009).
31 Rearing juvenile salmonids prefer shallow, relatively slow velocity habitat within or close
32 to cover such as LWM, inundated riparian vegetation, and submerged aquatic vegetation
33 (Beakes et al. 2014). During high flows, the shallow and slow velocity habitat with cover
34 is found on floodplains, in seasonal side channels, and other off channel habitat (Sommer
35 et al. 2001, Limm and Marchetti 2009). Shallow floodplains can also be very productive
36 resulting in fast growth rates for juvenile salmonids rearing on them (Sommer et al. 2001,
37 Jeffres et al. 2008, Katz et al. 2017). Channel incision resulting from substantially
38 diminished sediment supply reduces the availability of alternating bars and riffles as well
39 as side channels that juvenile Chinook salmon use for feeding and predator avoidance
40 during low flow periods (Beechie et al. 2005, Sellheim et al. 2015). During low flow
41 periods, side channels can provide the shallow, complex habitat that juvenile salmonids
42 prefer for rearing (Bellmore et al. 2013). Low water flows as a result of water regulation
43 are a major source of habitat degradation for native fishes in Reach 4B and the impacts of
44 low water flows are discussed in detail in the “Inadequate Flows” section below.

1 A separate but connected bypass system, consisting of the Chowchilla Bypass Channel,
2 Eastside Bypass Channel, and Mariposa Bypass Channel, was constructed to divert and
3 carry flood flows from the San Joaquin River and eastside tributaries upstream of the
4 Merced River (SJRRP 2010b). These bypasses are confined by manmade levees and have
5 limited floodplain access, habitat structure, nearshore habitat, and riparian habitat
6 required by Chinook salmon and other species.

7 Large quantities of downed trees are a functionally important component of many
8 streams (National Oceanic and Atmospheric Administration National Marine Fisheries
9 Service [NMFS] 1996, Beechie and Sibley 1997, Collins et al. 2002). Large woody
10 debris (LWD) influences channel morphology by affecting longitudinal profile, pool
11 formation, channel pattern and position, and channel geometry (SJRRP 2010b, Gurnell
12 et al. 2002). Downstream transport rates of sediment and organic matter are controlled in
13 part by storage of this material behind LWD. LWD also affects the formation and
14 distribution of habitat units, provides cover and complexity, and acts as a substrate for
15 biological activity (Collins et al. 2002, Roni et al. 2015). Wood enters streams inhabited
16 by salmonids, either directly from adjacent riparian zones, or from riparian zones in
17 adjacent nonfish-bearing tributaries (Latterell and Naiman 2007). Removal of riparian
18 vegetation and LWD from the streambank results in the loss of a primary source of
19 overhead and instream cover for juvenile salmonids. The removal of riparian vegetation
20 and LWD, and the replacement of natural bank substrates with rock revetment, can
21 adversely affect important ecosystem functions (Florsheim et al. 2008). Living space and
22 food for terrestrial and aquatic invertebrates is lost, eliminating an important food source
23 for juvenile salmonids. Loss of riparian vegetation and soft substrates reduces inputs of
24 organic material to the stream ecosystem in the form of leaves, detritus, and woody
25 debris, which can affect biological production at all trophic levels. The magnitude of
26 these effects depends on the degree to which riparian vegetation and natural substrates
27 are preserved or recovered during the life of the project.

28 Like many Central Valley rivers, the amount of LWD and potential recruitment into the
29 San Joaquin River below Friant Dam and specifically in Reach 4B has been severely
30 degraded by anthropogenic activities. Agricultural conversion, flood control, and water
31 development in addition to other anthropogenic activities have directly impacted the
32 LWD resources in Reach 4B. The riparian forests in Reach 4B as well as in upstream
33 reaches are substantially reduced from historical conditions (McBain and Trush 2002)
34 which has a direct impact on LWD. In addition, Friant Dam as well as smaller dams
35 downstream prevent or reduce the downstream movement of LWD.

36 Like LWD, shaded riverine aquatic habitat (SRA) is an important component of alluvial
37 river habitat for juvenile salmonids. SRA as defined by the USFWS (1992) is the
38 nearshore aquatic habitat occurring at the interface between a river and adjacent woody
39 riparian habitat and occurs from the edge of the bank to the limit of overhanging riparian
40 canopy or vegetation present within the water. The key attributes of SRA are: 1) the river
41 bank consisting of natural, eroding substrates supporting riparian vegetation that either
42 overhangs or protrudes into the water, and 2) the river containing varying amounts of
43 woody debris, often substantial detritus, and varying water velocities, depths, and flows
44 (USFWS 1992). Studies have demonstrated the importance of SRA in the Central Valley

1 for rearing juvenile salmonids with high juvenile salmonid densities associated with SRA
2 (USFWS 1998, USFWS 2010, USFWS 2011, Beakes et al. 2014).

3 The construction of levees and dikes to convert land for agricultural production tends to
4 channelize riverine habitats and reduces channel migration and avulsion (McBain and
5 Trush 2004). Reduced channel migration in the Restoration Area has eliminated off-
6 channel habitats, reduced complex side channels, and reduced instream habitat
7 complexity including large woody debris and riparian vegetation, particularly SRA,
8 which all serve to provide suitable conditions for juvenile salmonids over a wide range of
9 flows (SJRRP 2010b). Agricultural conversion also has directly reduced the amount of
10 floodplains, and levees and dikes have further isolated historic floodplains from the
11 channel. It is likely that the loss of floodplain habitats has substantially reduced food
12 resources and predator refuge for juvenile salmonids.

13 ***Hatchery Operations***

14 Seven hatcheries in the Central Valley raise anadromous salmonids, including in the
15 Sacramento River Basin, the Coleman National Fish Hatchery (Battle Creek), Feather
16 River Fish Hatchery, Nimbus Fish Hatchery (American River), Mokelumne River
17 Hatchery, Livingston Stone Hatchery (Sacramento River); and in the San Joaquin River
18 Basin, the: Merced River Fish Hatchery and the Salmon Conservation and Research
19 Facility (San Joaquin River) (McEwan 2001, SJRRP 2016). The Salmon Conservation
20 and Research Facility (SCARF), which is adjacent to the San Joaquin River below Friant
21 Dam, is being built to help meet the SJRRP goal of restoring self-sustaining runs of
22 Chinook Salmon (SJRRP 2016). SCARF is being designed and will be operated as a
23 conservation facility which will reduce or eliminate many of the negative impacts
24 associated with production/mitigation hatcheries. Hatchery production can negatively
25 affect fish populations by leading to a loss of genetic integrity primarily through
26 hybridization, inbreeding, and random genetic change (SJRRP 2010a). Hybridization
27 presumably creates individuals that are less well-adapted to local conditions than either
28 parent (Araki et al. 2008, Laikre et al. 2010). Inbreeding results from the breeding of
29 closely related individuals and is likely to develop from hatchery production because
30 eggs and milt are obtained from relatively few individuals (Wang et al. 2002). A small
31 breeding population also may lead to genetic drift. Both inbreeding and genetic drift can
32 lead to the production of individuals that are less adapted than naturally produced fish to
33 the natural environment in which the species evolved (Wang et al. 2002).

34 The following are other potentially negative effects of producing hatchery fish:

- 35 • Displacement of wild salmonid juveniles through competition and predation
36 (Levin et al. 2001, Tatara and Berejikian 2012)
- 37 • Competition between hatchery adults and wild adults for limited spawning habitat
38 (Kostow 2009)
- 39 • Stimulation of sport and/or commercial harvest efforts, which could increase the
40 harvest rate of naturally produced salmonids (NMFS 2016)

- 1 • Increase in disease rate among naturally produced fish (Miller et al. 2014)
- 2 • Negative social interaction between hatchery salmonids and wild salmonids
- 3 (Berejikian et al. 1996, Weber and Fausch 2005)

4 ***Impaired Water Quality***

5 **High Water Temperatures**

6 Release temperatures from Friant Dam under existing Friant operations typically
7 fluctuate between 48 to 58°F (8.9 to 14.4°C), and water temperatures are expected to be
8 suitable for juvenile rearing except in the downstream reaches (Reaches 2B to 5) as water
9 temperatures increase. However, release temperatures during recent drought years were
10 higher than typical with temperatures in the late summer through early winter ranging
11 into the upper 60s and low 70 degrees Fahrenheit (USGS gage 11251000 San Joaquin
12 River below Friant Dam). Critical to lethal water temperatures and exaggerated
13 fluctuations in water temperature result from a combination of factors, including
14 seasonally high air temperatures (May through September), low flow releases,
15 groundwater pumping that eliminated the inflow of cool groundwater throughout the
16 Restoration Area (thermal refugia), removal of large woody riparian forests that provided
17 shade, warm agricultural runoff, and warm flood flows from the Kings River through the
18 James Bypass (SJRRP 2010b). It is also possible that high flow releases during summer
19 and fall could exhaust the cold-water pool in Millerton Lake, thereby causing release
20 temperatures to substantially increase above 58°F (14.4°C). Many of these impacts would
21 directly affect the in-river life stages of anadromous salmonids. In the Fisheries
22 Management Plan, the SJRRP established monthly water temperature objectives for the
23 in-river life stages of Chinook salmon (SJRRP 2010b) based on water temperature
24 criteria presented in the U.S. Environmental Protection Agency's (EPA) Region 10
25 Guidance for Pacific Northwest State and Tribal Temperature Water Quality (EPA 2003),
26 Rich (2007) Impacts of Water Temperature on Fall-run Chinook Salmon (*Oncorhynchus*
27 *tshawytscha*) and Steelhead (*O. mykiss*) in the San Joaquin River System, and Pagliughi
28 (2008) Lower Mokelumne River Reach Specific Thermal Tolerance Criteria by Life
29 Stage for Fall-Run Chinook Salmon and Winter-Run Steelhead. The water temperature
30 objectives are summarized below for the adult migration, juvenile rearing, and juvenile
31 outmigration life stages that are relevant to the Reach 4B/ESB Project. Adult Chinook
32 salmon during migration start to experience stress from high water temperatures between
33 62.6 and 68°F (17 to 20°C), with lethal temperatures being greater than 68°F (20°C)
34 (SJRRP 2010b). However, the migration data collected by Strange (2010) suggest that the
35 lethal temperature is higher than this. Rearing and outmigrating juveniles start to
36 experience stress at water temperatures between 64.4 and 70°F (18 to 21.1°C), with the
37 prolonged exposure lethal temperature being greater than 75°F (23.9°C) (Pagliughi 2008,
38 SJRRP 2010b). Although floodplain-rearing temperatures can exceed 17 to 20°C, these,
39 floodplains can benefit growth given an adequate food supply, even in the presence of
40 stress inducing temperatures (Jeffres et al. 2008).

41 Water temperature is a primary limiting factor for natural steelhead production on many
42 Central Valley streams (NMFS 2009). Although many Central Valley dams provide
43 downstream water releases intended to benefit fall-run Chinook salmon, most do not
44 provide cool water temperatures for steelhead during summer and fall, especially during

1 extended droughts (Moyle et al. 2008). Many dams are not able to provide cool water
2 because they were not designed for deep-water reservoir releases or they lack adequate
3 cold-water pool storage (McEwan 2001). Where releases of cold water occur throughout
4 the summer, resident populations of trout often develop and remain, limiting anadromous
5 behavior (SJRRP 2011a, Sogard et al. 2012). The SJRRP did not establish monthly water
6 temperature objectives for steelhead like was done for Chinook Salmon (SJRRP 2010b).
7 General temperature guidelines for steelhead would be based on a DFG proposal to
8 assess temperature impairment (DFG 2007), EPA guidelines (EPA 2003), and a report on
9 temperature impacts on fall-run Chinook salmon and steelhead (Rich 2007).

10 Water temperature is a primary limiting factor for natural production of spring-run
11 Chinook salmon on Central Valley streams (NMFS 1999). Appropriate water temperature
12 regimes below many dams cannot be maintained at levels comparable to temperature
13 regimes that were achieved naturally in the upper watersheds that previously provided
14 holding, spawning, incubation, and rearing habitat (SJRRP 2011a).

15 **Contaminants**

16 Both natural and anthropogenic factors influence water quality within the San Joaquin
17 River (Quinn and Karkoski 1998). For instance, the Grasslands Basin is a hydrologic unit
18 situated west of the San Joaquin River that naturally drains the area between the Westlands
19 Water District and Highway 140 to the San Joaquin River. The Basin soils are naturally
20 high in salts and of low permeability (Quinn and Karkowski 1998). The low permeability
21 combined with water importation has resulted in a shallow groundwater table. To
22 maintain productivity, the installation of artificial drainage was necessary in low-lying
23 agricultural areas (Quinn and Karkowski 1998). Drainage from the southern part of the
24 basin (41,000 hectares) contains high concentrations of trace elements and soluble salts
25 that are harmful to fish and wildlife. The primary constituents of concern are salt, boron,
26 and selenium (Quinn and Karkoski 1998). Water quality in the valley floor of the San
27 Joaquin River Basin has been impaired because of contamination from a variety of other
28 sources, including 1) aquatic and terrestrial herbicide application, 2) urban and
29 agricultural pesticide application, 3) trace elements from industrial and agricultural
30 activities and those naturally present in soils, and 4) effluent from wastewater treatment
31 plants and livestock operations, particularly dairy farms (SJRRP 2010b). Point sources of
32 pollution originate from single identifiable sources, whereas nonpoint sources originate
33 from many different sources. Examples of nonpoint sources are agricultural runoff (e.g.,
34 excess fertilizers, herbicides, and pesticides) and urban stormwater containing oil, grease,
35 heavy metals, polycyclic aromatic hydrocarbons, and other organics (Central Valley
36 Regional Water Quality Control Board [RWQCB] 1998). Impervious surfaces (e.g.,
37 concrete) tend to reduce water infiltration and increase stormwater runoff (NMFS 1996).
38 Recent studies suggest that chronic or sublethal effects of contaminants may be subtle
39 and difficult to detect. For example, early experimental studies indicated that hatchery-
40 reared juvenile Chinook salmon exposed to undiluted agricultural subsurface drainwater
41 from the west side of the San Joaquin River had greater than 75 percent mortality,
42 whereas there were no chronic detrimental effects on the growth and survival of the study
43 fish exposed to agricultural return flows that were diluted by greater than or equal to 50
44 percent (Saiki et al. 1992). However, other studies suggest that juvenile fall-run Chinook
45 salmon died in the laboratory after eating selenium-contaminated invertebrates and prey

1 fish over a 90-day period that were collected from the San Joaquin River Basin (Beckon
2 2007).

3 A recent study has also indicated a serious potential risk of pesticides/insecticides/
4 fungicides to exposed early life stages of Chinook salmon and aquatic invertebrates in the
5 Central Valley (Viant et al. 2006). A large number of pesticides/insecticides/fungicides
6 have been detected by water quality sampling programs in the San Joaquin River Basin,
7 including aldrin, carbaryl, chlorpyrifos, diazinon, dieldrin, diuron, heptachlor, lindane,
8 malathion, metribuzin, and trifluralin (Domagalski et al. 2000). Most contaminant water
9 quality problems occur in the lower Restoration Area (Reaches 3 through 5) where water
10 quality is influenced by a lack of freshwater inflow with the majority of water being
11 imported from the Sacramento and San Joaquin River Delta (Delta) and by agricultural
12 drainage, particularly from Mud and Salt sloughs. Multi-year studies by Domagalski et
13 al. (2000) and others (Brown 1997, Panshin et al. 1998) assessed a wide array of
14 contaminants. The growing number of chemical pesticides/insecticides/fungicides found
15 in the San Joaquin Valley is too large to encompass in this review. Furthermore,
16 accurately quantifying risks of individual pesticides/insecticides/fungicides or
17 synergistic effects of multiple pesticides/insecticides/fungicides is not easily validated;
18 most studies rely on comparing contaminant levels (from biota or the environment) to
19 literature values, regional or national statistics, or suitable reference sites.

20 The San Joaquin-Tulare study unit (essentially the San Joaquin Valley) was among the
21 first basins chosen for the United States Geological Survey (USGS) National Water
22 Quality Assessment Program (NAWQA) and recently has focused considerable attention
23 on pesticide contamination in the San Joaquin River Basin (Dubrovsky et al. 1998,
24 Panshin et al. 1998, Kratzer and Shelton 1998, Brown and May 2000). Generally, toxicity
25 within the San Joaquin River has been attributed to pesticides/insecticides/fungicides from
26 agricultural nonpoint sources, substantiated by the lack of detection of pesticide compounds
27 in reference sites on the upper Kings River and Tuolumne River, situated above
28 agricultural influences (Dubrovsky et al. 1998). In the NAWQA studies, available
29 drinking water standards were not exceeded at San Joaquin River monitoring sites, but the
30 concentrations of several pesticides/insecticides/fungicides exceeded the criteria for the
31 protection of aquatic life. As mentioned previously, regional or national contamination
32 levels are used to interpret San Joaquin River study results. Gilliom and Clifton (1990,
33 from Brown 1998) reported that the San Joaquin River had some of the highest
34 concentrations of organochlorine residues in bed sediments among the major rivers of the
35 United States. Although the organochlorine pesticide DDT (dichloro-diphenyl-
36 trichloroethane) was banned in the United States in 1973, DDT concentrations have
37 continued to be detected in biota of the San Joaquin Valley streams at lower levels
38 (Goodbred et al. 1997, Dubrovsky et al. 1998), as contaminated soils are transported to
39 streams and sediment is resuspended from riverbeds. The most recent 303(d) list of
40 impaired waterbodies presented by the Central Valley RWQCB identifies Reaches 3, 4,
41 and 5 of the San Joaquin River study area, Mud Slough, and Salt Slough, all as impaired
42 due to pesticides and unknown toxicity.

43 Selenium and mercury are two environmental contaminants of primary concern in aquatic
44 environments, and the San Joaquin River is not an exception (SJRRP 2010b). Selenium

1 and mercury are trace elements that can be harmful to aquatic life because they
2 undergo biomagnification after being converted to organic forms in reducing (i.e., low
3 oxygen) conditions by methylating bacteria. Because of this conversion to an organo-
4 metallic compound, methylated selenium and mercury are absorbed preferentially into
5 fatty tissues and can biomagnify through the food chain despite low ambient
6 concentrations. Central Valley RWQCB water quality objectives for selenium are
7 currently being exceeded for Mud Slough and downstream reaches. While the reported
8 background concentrations for selenium for the San Joaquin River above Salt and Mud
9 sloughs are about 0.5 micrograms per liter ($\mu\text{g/L}$), selected sites along the river have
10 selenium concentrations from 1 to 5 $\mu\text{g/L}$ (Central Valley RWQCB 2001). The input of
11 selenium from the Grasslands area into the San Joaquin River represents a major risk for
12 larval fish, including Chinook salmon (Beckon 2007).

13 In past surveys, fish from several locations within the study area were shown to contain
14 elevated concentrations of arsenic (Rasmussen et al 1995, Saiki 1989), mercury (Rasmussen
15 et al. 1995, Saiki and May 1988), and selenium (Saiki 1989, Saiki and Lowe 1987, Saiki
16 and May 1988, White et al. 1988). A study by Saiki et al (1992), found that arsenic,
17 mercury, and selenium measured in composite whole-body samples of five fishes —
18 bluegill (*Lepomis macrochirus*), common carp (*Cyprinus carpio*), mosquitofish
19 (*Gambusia affinis*), largemouth bass (*Micropterus salmoides*), and Sacramento blackfish
20 (*Orthodon microlepidotus*) — from the San Joaquin River system were elevated;
21 however, only selenium approached concentrations that may adversely affect survival,
22 growth, or reproduction in warm water fishes. Moreover, only selenium among the four
23 measured elements exhibited a geographic (spatial) pattern that coincided with known
24 inflows of tile drainage to the San Joaquin River and its tributaries (Saiki et al. 1992).
25 Historical data from the Grassland Water District (a region exposed to concentrated tile
26 drainage) suggested that concentrations of selenium in fishes were at maximum during or
27 shortly after 1984, and have been slightly lower since then. The decline of selenium
28 concentrations in fishes from the Grasslands Water District area could be temporary if
29 additional acreages of irrigated lands in this portion of the San Joaquin Valley must be
30 tile-drained to protect agricultural crops from rising groundwater tables.

31 The 2010 SJRRP Annual Technical report presented water quality monitoring results for
32 compounds that could have potential effects on Chinook salmon and other fish native to
33 the San Joaquin River (SJRRP 2011c). Prominent findings included concentrations of
34 bifenthrin in sediment samples with the potential to cause mortality in certain organisms
35 and bioaccumulate up the food web and 30 water quality samples with copper exceeding
36 the United States Environmental Protection Agency (EPA) aquatic-life acute benchmark
37 for invertebrates.

38 ***Inadequate Flows***

39 Adult salmon passage below Friant Dam during the 1940s was inhibited by low flows in
40 the channel. In 1944 and 1947, California Department of Fish and Game (CDFG, now the
41 California Department of Fish and Wildlife [CDFW]) (1955a) observed 5,000 to 6,000
42 spring-run Chinook salmon migrating up the San Joaquin River as far as Mendota Dam
43 with flow that was estimated to be 100 cubic feet per second (cfs) in the reach between
44 Sack Dam and the confluence with the Merced River. CDFW (CDFG 1955a) observed

1 that many of these fish had rubbed themselves raw going over the shallow sandbars
2 between Sack Dam and the confluence with the Merced River (approximately 50 miles).
3 Such abrasions may increase the risk of mortality from disease for spring-run Chinook
4 salmon since they must spend an extended period of time holding in pools throughout the
5 summer before spawning in early fall (SJRRP 2010b). Abrasions on fish can increase the
6 probability of disease infection (Bader et al. 2006). Passage for the San Joaquin River
7 adult spring-run Chinook salmon has been blocked completely in the Restoration Area
8 since the 1950s when the river was dewatered below Sack Dam except during
9 uncontrolled flow releases in wet years (SJRRP 2010b).

10 Suitable flows are necessary year-round for juvenile salmon rearing. As flow increases,
11 the area preferred by juvenile Chinook salmon shifts from the center of the channel to
12 submerged terrestrial vegetation on the edge of the channel and within the floodplain
13 (SJRRP 2011b). Deeper inundation provides more overhead cover and protection from
14 avian and terrestrial predators than shallow water (Everest and Chapman 1972). In broad
15 low-gradient rivers, changes in flows can greatly increase or decrease the lateral area
16 available to juvenile Chinook salmon, particularly in riffles and shallow glides.

17 The Central Valley stream reaches (Mill, Deer, Butte creeks) that are presently accessible
18 to spring-run Chinook salmon often lack the summer habitat conditions needed to sustain
19 juvenile spring-run demonstrating the yearling life history in their lower reaches and
20 during drought years (SJRRP 2011b). These conditions can be exacerbated by reservoir
21 operations and water diversions that reduce summer flows and can be particularly severe
22 in drought years.

23 Reduced flows also interact with other stressors such as temperature, contaminants, other
24 water quality parameters, and disease to exacerbate conditions. A reduced volume of
25 water flow generally increases in temperature faster. Contaminants are less diluted in a
26 reduced flow volume. Disease transmission is increased when reduced flows reduce the
27 area of suitable habitat and cause fish to become more concentrated in the available
28 habitat.

29 ***Passage Impediments***

30 Fish migrate to spawn, feed, avoid predators, and escape stressful environmental
31 conditions. The success of migration, whether upstream, downstream or laterally (to
32 floodplain and off channel habitat) is limited by aquatic conditions and the presence of
33 barriers that can impede fish passage.

34 According to NMFS (2008), a passage impediment is defined as any artificial structural
35 feature or project operation that causes adult or juvenile fish to be injured, killed,
36 blocked, or delayed in migration, to a greater degree than in a natural river setting.

37 Direct and indirect impacts related to creating passage issues for migrating fish include:

- 38 • Blockage – Both complete and partial physical prevention of further migration.
39 Complete blockages prevent migration at all flow levels while partial blockages
40 only prevent migration at certain flow levels or only a portion of the fish are able

1 to pass (for example a blockage of a certain height that only 25% of Chinook
2 Salmon are able to jump over to continue their migration).

3 • Migration Delay- Opportunities to veer off course delaying migration, adding
4 stress, reducing energy stores, and potentially experiencing high temperatures

5 • Fatigue – Cannot complete immediate passage or reduces ability to complete
6 migration or life strategy

7 • Vulnerability – Predation and disease

8 • Injury – Impact, scrapes, and abrasions

9 • Desiccation – Tissue damage or reduction in gill function due to being out of
10 water for prolonged periods

11 • Disorientation – Fish cannot find pathway or access to passage, impeding or
12 reducing migration success

13 Velocity, depth, and elevational changes (hydraulic drops) can block or impede fish
14 movement. Whether a structure is an impediment to fish movement depends on the
15 physical and hydraulic features of the structure, and the physiology and behavior of the
16 fish; this can change with fish species and age. Barriers may create velocity, depth, and
17 slope conditions that fish cannot physically overcome, and these factors may disorient
18 fish or cause fish to avoid such conditions. In addition, turbulence, depth, and fall can
19 injure or otherwise incapacitate fish, increasing their vulnerability to predation, disease,
20 and fatigue. Multiple impediments along a migratory path may fatigue fish as they
21 migrate upstream or downstream and the cumulative effect of these impediments may
22 decrease the physical abilities of individual fish to migrate and successfully complete
23 their life history (Jones and Stokes 2001; Gallagher 1999).

24 In 2001, a fish passage evaluation of the entire Restoration Area classified potential
25 passage impediments as (Jones and Stokes 2001):

26 1. Entrainments

27 • Diversions/returns (diversions may or may not have a mechanical pump)

28 • Confluences

29 2. Barriers

30 • Structures (e.g., dams, headgates, control structures)

31 • Bridges

32 • Road crossings (e.g., mounded dirt, with or without culvert, spans channel)

33 The evaluation used past reports and documents, along with aerial photographs and
34 ground-truthing, to identify 90 potential impediments to migrating fish within Reach 4B.

1 A decade later, the California Department of Water Resources (DWR) performed another
2 fish passage evaluation for the SJRRP (SJRRP 2011b, 2012). This fish passage
3 evaluation was broken up into two tasks. Task 1 was an initial evaluation of structures in
4 the Restoration Area and included identification and data collection of potential fish
5 passage barriers, identification of fish passage criteria for the evaluation, and
6 identification of potential barriers for future study (SJRRP 2011b). Task 2 consisted of
7 data collection and hydraulic evaluation of the potential fish passage barriers identified in
8 Task 1 (SJRRP 2012).

9 Task 1 only evaluated structures that would have an impact on migration of fish in the
10 San Joaquin River and bypasses, and did not consider off-channel structures such as
11 diversions or gravel mining pits (SJRRP 2011b). DWR reviewed existing reports on
12 potential fish passage barriers, which resulted in the identification of 61 structures that
13 were reviewed for inclusion in the Task 1 evaluation. Of these 61 structures, only 18 were
14 included for analysis in Task 2 based on the Task 1 criteria. In addition to previously
15 identified structures, DWR identified other potential fish passage barriers through
16 examination of aerial photographs, and built structural models in the San Joaquin
17 Hydrologic Engineering Center River Analysis System (HEC-RAS), as part of the Task 1
18 analyses. These analyses resulted in the identification of 50 new structures for a total of
19 68 structures that were evaluated in Task 1. A total of 45 of the 68 structures were field
20 surveyed using the First Pass method, with the remaining structures not surveyed due to
21 access issues (SJRRP 2011b). The First Pass method consisted of physical data collection
22 of each structure including measurements and photographs (SJRRP 2011b). An
23 additional structure was added based on the field observation and several more structures
24 were evaluated based on existing field data or data collected at a distance (SJRRP
25 2011b). The data from the First Pass survey were analyzed with ArcGIS GeoDatabase
26 using the fish passage criteria identified for use in this analysis (SJRRP 2011b). The
27 ArcGIS GeoDatabase categorized the structures as Green (not a barrier to fish migration
28 and will not be further analyzed), Gray (placed on a list for Second Pass analysis), and
29 Red (fish passage barrier). This resulted in the identification of 28 structures that were
30 Green, 13 as Gray, and 8 as Red (SJRRP 2011b).

31 In Task 2, the 13 structures identified as potential fish passage barriers (Gray) in Task 1,
32 as well as two Red barriers (Eastside Bypass and Mariposa Bypass control structures),
33 were evaluated using data collection and hydraulic evaluation (SJRRP 2012). In addition,
34 the Merced NWR weirs in the Eastside Bypass were identified as potential barriers by
35 United States Department of the Interior, Bureau of Reclamation (Reclamation) and
36 evaluated in Task 2. In evaluating fish passage for Task 2, criteria were identified based
37 on guidelines developed by CDFW, NMFS, and others for adult salmonids (SJRRP 2011b,
38 SJRRP 2012). Due to the complexity of developing criteria and evaluating every structure
39 for all fish species potentially present in the reach, adult Chinook salmon were selected
40 as the focal species of the evaluation (SJRRP 2012). However, the SJRRP Native Fish
41 Attributes Table with fish passage criteria was also considered (SJRRP 2012). Fish
42 passage at all identified structures was evaluated based on three main criteria: jump
43 height into the structure, depth in the structure, and velocity in the structure. Second Pass
44 data collection was focused on the data needed to create hydraulic models for the sites
45 and included flow, velocities, and depth. Hydraulic data and models were evaluated in

1 relation to fish capabilities in order to determine Chinook salmon passage success at each
2 potential San Joaquin River fish barrier. The flow ranges used in the model for fish
3 passage was 25 – 4,500 cfs for the San Joaquin River and 25-8,500 cfs for the bypasses
4 with the flow being the actual flow at the structure and not the release from Friant
5 (SJRRP 2012).

6 **Entrainment**

7 Herren and Kawasaki (2001) found 298 diversions in the San Joaquin River Basin. More
8 than 95 percent of these diversions were unscreened at the time of the study with
9 unscreened diversions increasing the likelihood of fish entrainment. The precise impacts
10 of these diversions across life stages of Chinook salmon or other fishes are unknown
11 (SJRRP 2010a). No studies have been conducted to determine the entrainment rates at
12 pumps and weirs within the Restoration Area (SJRRP 2010a). In a laboratory experiment,
13 smolt sized juvenile Chinook Salmon were found to have an entrainment risk of 0.3 to
14 2.3 percent when encountering a simulated unscreened water diversion (Mussen et al.
15 2013). In a juvenile Chinook Salmon entrainment study of agricultural pumps in the
16 Sacramento River, an average of 0.05 percent (range 0 to 1.0 percent) of marked salmon
17 released upstream of the diversion were recaptured (Hanson 2001).
18 Water diversions can reduce survival of emigrating juvenile salmonids by causing direct
19 losses at unscreened or inadequately screened diversions; these diversions can also cause
20 indirect losses associated with reduced streamflows (SJRRP 2010a). Fish screening and
21 salvage efforts at major agricultural diversions have met with variable levels of success,
22 and many smaller unscreened or inadequately screened diversions continue to operate.
23 Unscreened diversions continue to be operated due to the lengthy fish screen regulatory
24 permitting process and they can be expensive to install. Fish losses at diversions can
25 result from physical injury, impingement, entrainment, or predation. Delayed passage,
26 increased stress, and increased vulnerability to predation also contribute to mortality
27 caused by diversions. Diversion impacts on migratory fish depend on diversion timing
28 and magnitude, river discharge, fish species and life stage, and other factors.

29 *Diversions/returns (Diversions may or may not have a mechanical pump)* Sixty
30 diversion/returns within the 4B Reach were identified using aerial photographs (Jones
31 and Stokes 2001). Three of those locations were visited by the 4B Fisheries Team in
32 2010. One location was a pipe culvert with a flap gate, and the other two locations were
33 pumping stations. These facilities appeared to have the potential to entrain juvenile fish.
34 At all three locations, culvert configurations indicate that once fish were diverted from
35 the channel, they could not return (Figure G-1). Debris screening was observed at two of
36 the three locations, providing a barrier to adult fish, but could entrain most sizes of
37 juvenile fish. All three locations had large cut ditches from the channel to the diversion or
38 return. These areas were relatively deep, and had low flow velocity, indicating a potential
39 for harboring piscivorous predators.



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Figure G-1.
Example of Pump Diversion within Reach 4B

4 *Tributary Confluences (False Pathways)* using aerial photographs, five confluence
5 connections were identified within Reach 4B with the main confluences being with Bear
6 and Owens creeks, and all were located on the Eastside Bypass (Jones and Stokes 2001).
7 During high flows, there could be the potential to attract migrating adult fish from the main
8 channel, which could create negative consequences like migration delays, missed cues, or
9 exposure to elevated temperatures. Juveniles might also traverse these tributaries, but this
10 may or may not have negative consequences, depending on whether these areas provide
11 beneficial rearing habitat, and if juveniles could freely return to the main channel.

12 **Barriers** As described above, DWR performed a fish passage evaluation for the SJRRP
13 throughout the Restoration Area (SJRRP 2011b, 2012b). In evaluating fish passage,
14 criteria were selected based on guidelines developed by CDFW, NMFS, and others for
15 adult salmonids (SJRRP 2011b, SJRRP 2012). Due to the complexity of developing criteria
16 and evaluating every structure for all fish species that may be present in the reach, adult
17 Chinook salmon was the focus species of the evaluation but passage for all native fish
18 species was considered (SJRRP 2012). Fish passage potential at all identified structures,
19 at flows ranging from 25 to 4,500 cfs at the potential barriers in the San Joaquin River
20 and 25 to 8,500 cfs for potential barriers in the bypass, was evaluated based on three main
21 criteria: jump height into the structure, water depth within the structure, and flow velocity
22 within the structure. Hydraulic data were evaluated in relation to fish capabilities in order
23 to determine Chinook salmon passage success at each potential San Joaquin River fish
24 barrier.

25 The results of the Task 2 evaluation conducted by DWR, suggested that adult Chinook
26 salmon would not be able to pass structures at most flows in Reach 4B, or the Eastside

1 Bypass, unless improvements are completed to allow passage (SJRRP 2012). The
2 following eight structures in the Reach 4B/ESB Project study area were identified as
3 either partial or complete barriers for adult migration of salmon and would be evaluated
4 further to develop passage alternatives (SJRRP 2012):

- 5 • Merced Refuge Weir #2
- 6 • Merced Refuge Weir #1
- 7 • Dan McNamara Road
- 8 • Eastside Bypass Control Structure
- 9 • Mariposa Bypass Control Structure
- 10 • Mariposa Drop Structure
- 11 • Eastside Bypass Rock Weir

12 The restriction of spawning to a limited area below impassable barrier is considered one
13 of the primary factors that explains the decline of Central Valley anadromous fish
14 species, including Chinook salmon and steelhead (SJRRP 2010a). Barriers can also
15 impede the movement of numerous other native and non-native fish species.

16 **Flow Structures**

17 *Reach 4B Headgates* The Headgates are located at RM 168. They consist of an earth fill
18 dam with four, square concrete headgate culverts controlling flow into Reach 4B. When
19 the gates are closed, this structure is a complete barrier to flow and fish. The gates have
20 not been operational for many years (and may no longer be operational) but would be a
21 fish passage barrier if they could be operated (Figure G- G-2). If the gates could be
22 opened the structure would require consistent maintenance due to the small diameter of
23 each culvert and there is a high probability that the culverts will become plugged with
24 debris.



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**Figure G-2.
Four Culverts Associated with the Reach 4B Headgates**

4 There also appears to be an elevation gradient that would be an impediment to upstream
 5 and downstream migration. The structure also would have debris load issues that would
 6 further impede fish movement. Energy dissipation would create a potential pool in
 7 conjunction with the concrete basin, providing holding areas for predators of small fish
 8 moving downstream. Depending on velocities, fish might impact concrete energy
 9 dissipation structures, causing injury or disorientation. High concentrations of invasive
 10 aquatic vegetation could potentially influence water quality (i.e., dissolved oxygen)
 11 adjacent to the structure, creating an additional physiochemical barrier for some fish.

12 *Sand Slough Control Structure* Located adjacent to the Reach 4B Headgates is the Sand
 13 Slough Control Structure (RM 168). This is a low head control structure in Sand Slough
 14 between the San Joaquin River and the Eastside Bypass (Figure G-3). Task 2 determined
 15 that this structure is not a fish barrier (SJRRP 2012). However, the large scour pools
 16 above and below the concrete structure could provide potential predator holding areas.
 17 Predation on juvenile salmon can be quite high within energy dissipation pools located
 18 below control structures (Sabal et al. 2016).



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**Figure G-3.
Sand Slough Control Structure**

4 *Mariposa Bypass Control Structure* This structure is located at ~RM147 within the
5 Mariposa Bypass. The concrete has 14 bays (6 open in the middle and 4 gated on either
6 side; Figure G-4). Each of the bays has concrete energy dissipation structures that would
7 create upstream fish barriers under a variety of flows. The Mariposa Bypass Control
8 Structure is a barrier at all flows (SJRRP 2012). Manipulation of the gates would likely
9 not improve passage. Dissipation structures most likely would create hydraulic drops that
10 could potentially injure and disorient downstream moving fish. A combination of scour
11 holes and dissipation sills could create stranding and predation issues for juvenile fish. At
12 lower flows, the pool just downstream of the structure would greatly dissipate velocities,
13 creating an energy sink for juvenile fish and potentially disorient fish searching for
14 upstream and downstream passage. This pool also might create water quality issues,
15 including temperature and dissolved oxygen barriers as well as elevated risk of predator
16 holding.



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**Figure G-4.
Mariposa Bypass Control Structure**

4 *Mariposa Bypass Drop Structure* This structure is located at ~ RM147 in the Eastside
 5 Bypass and diverts flow from the Eastside Bypass to the Mariposa Bypass. The structure
 6 consists of a concrete wall spanning the channel and two concrete walls framing the
 7 downstream channel. The channel-spanning wall is over 6 feet tall on the upstream side
 8 and well over 15 feet on the downstream side. The wall is likely a barrier at all flows
 9 even when completely inundated during flood flows. The concrete basin on the
 10 downstream side concentrates high flows, creating a very large scour pool (well over
 11 1 acre in size). At lower flows, this pool would greatly dissipate velocities, creating an
 12 energy sink for juvenile fish and potentially disorient fish searching for upstream and
 13 downstream passage and create an elevated risk of predator holding (Figure G-5).



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Figure G-5.
Scour Hole Associated with the Mariposa Bypass Drop Structure

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The downstream hole could also create potential water quality issues during lower flow situations (e.g., temperature and dissolved oxygen). Scour holes at the top and bottom of the structure could create potential predator holding areas.

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Eastside Bypass Control Structure This structure is in the Eastside Bypass immediately adjacent to the Mariposa Bypass Bifurcation Structure. The structure constricts flow through six radial gates. Each of the bays has concrete energy dissipation structures that would create upstream fish barriers under a variety of flows (see Figure G-6). The energy dissipating blocks create physical passage barriers to large fish (i.e., adult anadromous salmonids). There are weep holes, small holes designed for water release at low flows (see Figure G-6), across the wall face, but their utility in passing fish appears minimal. Manipulation of the gates might improve some passage but may also cause potential impingement issues. Structures most likely would create hydraulic drops that could potentially injure and disorient downstream moving fish.

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**Figure G-6.
Energy Dissipation Sills, Radial Arms, and Weep Holes within
the Eastside Bypass Control Structure**

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A combination of scour holes and dissipation sills could create stranding and predation issues for juvenile fish. At lower flows, the lower pool would greatly dissipate velocities, creating an energy sink for juvenile fish and potentially disorient fish searching for upstream and downstream passage. This pool also might create water quality issues, including temperature and dissolved oxygen barriers.

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National Wildlife Refuge Weirs Within the Eastside Bypass, two low weirs control water elevation and flow in the wildlife refuge. Both structures appear to create upstream and downstream barriers to fish due to hydraulic drops. Passage would be further impeded due to high debris loading across both structures from plant production and beaver (*Castor canadensis*) activity (Figure G-7). Predation could be enhanced because of low velocities in and around constricted passage areas.

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Figure G-7.
Low Head Weir within the Wildlife Refuge

4 *Other Potential Barriers*

5 *Bridges* Nine bridges were identified based on aerial photographs of Reach 4B
6 (Jones and Stokes 2001). The bridges do not appear to create any major passage issues.
7 However, high concentrations of bridge abutments could collect debris during high flows,
8 causing backwater conditions and creating passage issues (Figure G-8). Bridges
9 constructed with concrete aprons or energy dissipation structures may create depth and
10 velocity barriers at low flows and scour holes downstream of the structures that could
11 block fish movement. Any blockages that reduce flow velocity or create scour holes
12 could generate conditions advantageous to predators.



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Figure G-8.
Example of Bridge and Abutments within Reach 4B

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Road crossings (mounded dirt; with or without culvert; spans channel)

5 Seven road crossings were identified based on a review of aerial photographs (Jones and
 6 Stokes 2001). Two of these crossings were visited in 2010 by the 4B Fisheries Team.
 7 Both crossings were earthen mounds spanning the entire San Joaquin River channel, with
 8 a single, corrugated pipe culvert passing through each. The culverts were significantly
 9 under-sized for the channel and would not be able to carry the range of flows expected.
 10 Both culverts would have debris loading issues, and the crossings most likely would dam
 11 water and then overtop under most flows (Figure G-9). Potential debris and depth barriers
 12 are created under this situation. Upstream migrating fish would not be able to negotiate
 13 these culverts. If kept clean, downstream passage of some smaller fish (e.g., juvenile
 14 salmonids) would be possible. Elevated earthen mounds and undersized pipes most likely
 15 would create scour on the downstream side of crossings, creating potential predator
 16 holding areas and hydraulic drop barriers under most situations. Some of these road
 17 crossings may wash out during high flows (McBain and Trush 2002). The seven
 18 crossings identified in 2001 may not be existing currently, particularly after the high
 19 flows of 2017.



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Figure G-9.
Example of Road Crossing and Associated Culvert

4 **Predation**

5 San Joaquin River fish assemblage monitoring conducted during 2003 to 2005 (CDFG
6 2007) and 2012 to 2014 (SJRRP 2014, 2017) indicated that the Reach 4B/ESB Project
7 area is inhabited by several non-native species that are known to prey on juvenile
8 salmonids and other native species, including largemouth bass (*Micropterus salmoides*),
9 green sunfish (*Lepomis cyanellus*), warmouth (*Lepomis gulosus*), black crappie (*Pomoxis*
10 *nigromaculatus*), white crappie (*Pomoxis annularis*), striped bass (*Morone saxatilis*),
11 bluegill (*Lepomis macrochirus*), pumpkinseed (*Lepomis gibbosus*), redear sunfish
12 (*Lepomis microlophus*), and spotted bass (*Micropterus punctulatus*) (Grossman 2016).

13 There is an apparent shift in species composition from native to non-native fish
14 assemblages (predominated by predator centrarchid species) with increasing distance
15 downstream from Friant Dam (CDFG 2007, SJRRP 2014, 2017). There was also a
16 corresponding downstream shift in habitat type dominance by area, from glides to pools.
17 In California streams, some species, such as introduced centrarchids, tend to increase
18 their populations with increased human disturbance of habitats, including lowered stream
19 flows, increased number of pools, and increased turbidity (Moyle and Nichols 1973). The
20 more downstream reaches (e.g., the Reach 4B/ESB Project area) were completely
21 dominated by non-native species (including many predator species) in all habitat types
22 (CDFG 2007).

23 High predation rates on migratory fish, including juvenile salmonids, have been observed
24 below small dams in Central Valley rivers (Tucker et al. 1998, Sabal et al. 2016). As

1 juvenile salmon pass over small dams, the fish are subject to conditions that may
2 disorient them, making them highly susceptible to predation by other fish or birds
3 (Beamesderfer et al. 1996, Wiese et al. 2008). In addition, deep pool habitats tend to form
4 immediately downstream from these dams where Sacramento pikeminnow, striped bass,
5 and other potential predators congregate (Sabal et al. 2016). Tucker et al. (1998) showed
6 high rates of predation by Sacramento pikeminnow and striped bass on juvenile salmon
7 below the RBDD, and Sabal et al. (2016) demonstrated high predation rates on
8 emigrating salmonids below Woodbridge Dam on the Mokelumne River by striped bass.

9 Striped bass, an invasive non-native anadromous species, which primarily migrate into
10 the San Joaquin River tributaries during the late-winter and spring (S.P. Cramer and
11 Associates 2004, 2005; Cramer Fish Sciences 2006, 2007), were the primary predators of
12 juvenile fall-run Chinook salmon fitted with radio tags in a Stanislaus River study
13 (Demko et al. 1998). Although more than 90 percent of the radio-tagged fish appear to
14 have been eaten by predators, there is uncertainty as to whether gastrically implanting the
15 radio tags, which had 12-inch-long external whip antennas, impaired the ability of the
16 juvenile salmon to avoid predators (Demko et al. 1998). A recent predation study in the
17 lower San Joaquin River found a mean relative predation rate of 15.3 percent on tethered
18 juvenile Chinook Salmon (Demetras et al. 2016). Of the 12 video documented predation
19 events, 3 were confirmed to be by striped bass with the other predators not identifiable to
20 species (Demetras et al. 2016).

21 Birds are also known to prey on juvenile salmonids and other fish species (Evans et al.
22 2012). Caspian terns *Hydroprogne caspia* were documented to prey on juvenile fall-run
23 Chinook salmon migrating in San Francisco Bay (Evans et al. 2011). In addition to terns,
24 double-crested cormorants *Phalacrocorax auritus*, California gull *Larus californicus*,
25 ring-billed gull *L. delawarensis*, and American white pelicans *Pelecanus erythrohynchos*
26 were documented to consume salmonids in the Columbia River basin (Evans et al. 2012).
27 The minimum predation rate of terns and cormorants on Willamete River spring-run
28 Chinook salmon was 2.5% (Evans et al. 2012). Western gulls *Larus occidentalis*
29 consumed juvenile coho salmon *Oncorhynchus kisutch* and steelhead in Central
30 California streams with some of the streams appearing to have high predation rates
31 (Osterback et al. 2013). To date, no avian predation studies on Central Valley salmon
32 have been performed.

33 **G.1.4 Fish Species**

34 Fish communities in the San Joaquin Reach 4B/ESB Project study area have changed
35 markedly in the last 150 years (SJRRP 2011b). Native fish assemblages were adapted to
36 widely fluctuating riverine conditions, ranging from large winter and spring floods to low
37 summer flows, and had migratory access to extensive upstream habitats. These
38 environmental conditions resulted in a broad diversity of fish species, including
39 anadromous species. Fishes that may have historically occurred, as well as those that
40 currently inhabit the Reach 4B/ESB Project area, are listed in Table G-1.

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Table G-1.
Fish species with historic or current presence within the
Reach 4B/ESB Project study area

Category	Species	Scientific Name	Federal/State Status¹	Current Presence
Native Anadromous	Central Valley Spring-run Chinook Salmon	<i>Oncorhynchus tshawytscha</i>	T/T	Periodic ²
	Central Valley Fall-run Chinook Salmon	<i>Oncorhynchus tshawytscha</i>	SC-/ SC	Periodic
	California Central Valley steelhead	<i>Oncorhynchus mykiss</i>	T/SC	Unknown; Rainbow trout observed in Reach 1
	North American Green Sturgeon	<i>Acipenser medirostris</i>	T/SC	No; Only anecdotal evidence of historic presence in San Joaquin River
	White Sturgeon	<i>Acipenser transmontanus</i>	--/SC	Yes ³ ; Observed by DIDSON in Reach 5
	River Lamprey	<i>Lampetra ayersii</i>	--/SC	Unknown; have not been observed in Restoration Area during surveys
	Pacific Lamprey	<i>Entosphenus tridentata</i>	--/SC	Periodic/ observed in Reach 1
Native Riverine	Sacramento Hitch	<i>Lavinia exilicauda exilicauda</i>	--/SC	No; Observed in Reach 2,3, and 5
	Sacramento Blackfish	<i>Orthodon microlepidotus</i>		Yes
	Sacramento Splittail	<i>Pogonichthys macrolepidotus</i>	--/SC	Periodic
	Sacramento Perch	<i>Archoplites interruptus</i>	--/SC	Extirpated
	Hardhead	<i>Mylopharodon conocephalus</i>	--/SC	No; Observed in Reach 1
	Sacramento Pikeminnow	<i>Ptychocheilus grandis</i>		No; Observed in Reach 1
	Sacramento Sucker	<i>Catostomus occidentalis occidentalis</i>		Yes
	Tule Perch	<i>Hysterocarpus traski</i>		No; Observed in Reaches 2 and 3
	Prickly Sculpin	<i>Cottus asper</i>		Yes
	Riffle Sculpin	<i>Cottus gulosus</i>	--/SC	No; Observed in Reaches 1 and 3
	Threespine Stickleback	<i>Gasterosteus aculeatus</i>		No; Observed in Reach 1
Native Resident Lamprey	Kern Brook Lamprey	<i>Lampetra hubbsi</i>	--/SC	No; Observed in Reach 1

Category	Species	Scientific Name	Federal/State Status ¹	Current Presence
Non-native Invasive Anadromous	Striped Bass	<i>Morone saxatilis</i>		Yes
Non-native Invasive Resident	Black Bullhead	<i>Ameiurus melas</i>		Yes
	Brown Bullhead	<i>Ameiurus nebulosus</i>		Yes
	Channel Catfish	<i>Ictalurus punctatus</i>		Yes
	White Catfish	<i>Ameiurus catus</i>		Yes
	Bigscale Logperch	<i>Percina macrolepida</i>		Yes
	Fathead Minnow	<i>Pimephelas promelas</i>		Yes
	Inland Silverside	<i>Menidia beryllina</i>		Yes
	Red Shiner	<i>Cyprinella lutrensis</i>		Yes
	Golden Shiner	<i>Notemigonus crysoleucas</i>		Yes
	Goldfish	<i>Carassius auratus</i>		
	Western Mosquitofish	<i>Gambusia affinis</i>		Yes
	Common Carp	<i>Cyprinus carpio</i>		Yes
	Shimofuri Goby	<i>Tridentiger bifasciatus</i>		Yes
	Black Crappie	<i>Pomoxis nigromaculatus</i>		Yes
	White Crappie	<i>Pomoxis annularis</i>		Yes
	Bluegill	<i>Lepomis macrochirus</i>		Yes
	Green Sunfish	<i>Lepomis cyanellus</i>		Yes
	Pumpkinseed	<i>Lepomis gibbosus</i>		No; observed in Reach 5
	Redear Sunfish	<i>Lepomis microlophus</i>		Yes
	Warmouth	<i>Lepomis gulosus</i>		Yes
	Spotted Bass	<i>Micropterus punctulatus</i>		Yes
	Largemouth Bass	<i>Micropterus salmoides</i>		Yes
	Threadfin Shad	<i>Dorosoma petenense</i>		Yes

Source: Fish presence information is from San Joaquin River fish assemblage monitoring conducted during 2012 to 2014 (SJRRP 2014, 2017).

¹ SC = California Species of Special Concern, T = Threatened

² CV Spring-run Chinook Salmon are a focus of SJRRP reintroduction activities

³ CDFG report card data 2009

- 1 The following species descriptions include a brief account of the current and historical
- 2 distribution, life history patterns, and habitat requirements of fish species with historic or
- 3 current presence in the Reach 4B/ESB Project area. This section is subdivided into native
- 4 anadromous fish, native riverine fish, non-native invasive/introduced anadromous
- 5 species, and non-native invasive/introduced riverine species.

1 **Native Anadromous Fish Species**

2 Due to the numerous fish barriers present in the Reach 4B/ESB Project area and lack of
3 adequate flows (see Stressors section), native anadromous fish species historically
4 present in the Restoration Area cannot access the Reach 4B/ESB Project area and reaches
5 upstream except in the wettest years. Therefore, all anadromous fish species effectively
6 have been extirpated from the Restoration Area because rare and inconsistent access has
7 not allowed viable populations to persist. Furthermore, extreme habitat degradation and
8 unsuitably high-water temperatures (see Stressors section) has made aquatic habitat in the
9 Restoration Area unsuitable for most life stages of native anadromous fish species. Since
10 there is only anecdotal evidence of the historical use of the San Joaquin River by North
11 American green sturgeon (Beamesderfer et al. 2004, Jackson and Van Eenennaam 2013)
12 this species is not further discussed in this document.

13 **Central Valley Spring-run Chinook Salmon** Spring-run Chinook salmon in the
14 Central Valley was once among the largest runs on the Pacific Coast (Yoshiyama et al.
15 1998). Construction of dams on the Sacramento, American, Mokelumne, Stanislaus,
16 Tuolumne, Merced, and San Joaquin rivers helped lead to the extirpation of spring-run
17 Chinook salmon from these watersheds. Annual abundance estimates of extant Central
18 Valley spring-run Chinook salmon populations display a high level of fluctuation but the
19 overall number of spring-run Chinook salmon remain far below estimates of historic
20 abundance (SJRRP 2011a). On September 16, 1999, NMFS listed the Central Valley
21 spring-run Chinook salmon evolutionarily significant unit (ESU) as threatened under the
22 Federal Endangered Species Act (ESA).

23 On January 30, 2014, NMFS designated a nonessential experimental population (NEP) of
24 Central Valley spring-run Chinook salmon under section 10(j) of the Endangered Species
25 Act in portions of the San Joaquin River below Friant Dam (78 FR 79622). The
26 experimental population area includes the San Joaquin River just upstream from its
27 confluence with the Merced River to Friant Dam including all sloughs, channels,
28 floodways, and waterways that CV spring-run Chinook salmon can access along the San
29 Joaquin River as well as portions of the Kings River when it is connected to the San
30 Joaquin River (high water years; 78 FR 79622). The NEP is treated under section 7 as if
31 it is a species proposed for listing. Protective regulations under section 4(d) and 9 apply
32 to the NEP within the experimental population area. The unintentional take of CV spring-
33 run Chinook Salmon in the experimental population area that is caused by otherwise
34 lawful activities is excepted from Section 9 take provisions. Outside of the experimental
35 population area, CV spring-run Chinook salmon will continue to be covered by the take
36 prohibitions and exceptions applicable to the non-experimental part of the ESU but
37 limited take exceptions will apply to meet the *de minimis* conditions of the settlement
38 (78 FR 79622). In the lower San Joaquin River and its tributaries downstream from the
39 Merced River confluence to Mossdale County Park, take of CV spring-run Chinook
40 salmon is excepted if the avoidance of such take would impose more than *de minimis*
41 impact on water supply reductions, additional storage releases, or bypass flows on
42 unwilling third parties (78 FR 79622).

43 In the San Joaquin River, spring-run Chinook salmon (*Oncorhynchus tshawytscha*)
44 historically spawned as far upstream as the present site of Mammoth Pool Reservoir (RM

1 322), where their upstream migration historically was blocked by a natural velocity
2 barrier (P. Bartholomew, pers. comm., as cited in Yoshiyama et al. 1996). The San
3 Joaquin River historically supported large runs of spring-run Chinook salmon; (CDFG
4 1990, as cited in Yoshiyama et al. 1996) suggested that this run was one of the largest
5 Chinook salmon runs on any river on the Pacific Coast, with an annual escapement
6 averaging 200,000 to 500,000 adult spawners (CDFG 1990, as cited in Yoshiyama et al.
7 1996). Construction of Friant Dam began in 1939 and was completed in 1942, which
8 blocked access to upstream habitat (SJRRP 2011b). Nevertheless, runs of 30,000 to
9 56,000 spring-run Chinook salmon were reported in the years after Friant Dam was
10 constructed, with salmon holding in the pools and spawning in riffles downstream from
11 the dam. Friant Dam began filling in 1944 and, in the late 1940s, began to divert
12 increasing amounts of water into canals to support agriculture. Flows into the main stem
13 San Joaquin River were reduced to a point that the river ran dry near Gravelly Ford. By
14 1950, the entire run of spring-run Chinook salmon was extirpated from the San Joaquin
15 River (Fry 1961).

16 Adult spring-run Chinook salmon historically used the Reach 4B/ESB Project area as a
17 migration corridor during upstream migration in early spring on their way to holding
18 habitat in the upper reaches of the San Joaquin River (Clark 1943). Historical migration
19 through the project area was April through June, with May being the peak time period.
20 Adult migration requires sufficient water depths and velocities to provide barrier-free
21 passage to holding and spawning habitat. CDFW uses a minimum depth of 0.9 feet for
22 passage of adult Chinook salmon in their passage assessments (CDFW 2016). Boles
23 (1988) recommends water temperatures below 65°F (18.3°C) for adult Chinook salmon
24 migration. Lindley et al. (2004) report that adult migration is blocked when temperatures
25 reach 70°F (21.1°C), and fish can become stressed as temperatures approach 70°F
26 (21.1°C). In contrast, Strange (2010) found that adult salmon migration in the Klamath
27 River was blocked at temperatures above about 73°F (22.8°C), with some migration
28 occurring up to temperatures of 75°F (23.9°C).

29 Spring-run Chinook salmon enter freshwater as sexually immature adult fish, and their
30 holding period can last for several months before individuals are ready to spawn in the
31 fall (Moyle 2002; CDFG 1998). Spring-run Chinook salmon historically spawned in
32 the San Joaquin River upstream from the town of Friant from late August to October,
33 peaking in September and October (Clark 1943). Egg incubation generally lasts
34 between 40 and 90 days at water temperatures of 43 to 54°F (6 to 12°C) (Vernier 1969,
35 Bams 1970, Heming 1982, Bjornn and Reiser 1991). Alevins remain in the gravel for 2 to
36 3 weeks after hatching and absorb their yolk sac before emerging from the gravels into
37 the water column from November to March (Fisher 1994, Ward et al. 2003).

38 The length of time spent rearing in freshwater varies greatly among juvenile spring-run
39 Chinook salmon across their range (SJRRP 2011b). Spring-run Chinook salmon may
40 disperse downstream as fry soon after emergence, early in their first summer, in the fall
41 as flows increase, or as yearlings during the spring after overwintering in freshwater
42 (Healey 1991). In contrast to more northern spring-run Chinook salmon populations,
43 many of the current Central Valley populations exhibit fry and smolt downstream
44 migration during the winter and spring of their first year, and relatively few exhibit a

1 yearling life history (NMFS 2014). However, some juveniles likely migrate downstream
2 throughout the year (Nicholas and Hankin 1989).

3 Historically, spring-run Chinook salmon juveniles likely used the Reach 4B/ESB Project
4 area as a migration corridor and rearing area due to the extensive floodplain habitat
5 present. Juvenile salmonids rear on seasonally inundated floodplains when available.
6 Sommer et al. (2001) found higher growth and survival rates of Chinook salmon
7 juveniles reared on the Yolo Bypass compared with those in the main stem Sacramento
8 River. Jeffres et al. (2008) observed similar results on the Cosumnes River floodplain.
9 Drifting invertebrates, the primary prey of juvenile salmonids, were more abundant on
10 the inundated Yolo Bypass floodplain than in the adjacent Sacramento River (Sommer et
11 al. 2001).

12 **Central Valley Fall-run Chinook Salmon** Fall-run Chinook salmon generally spawned
13 lower in the watershed than spring-run Chinook salmon (CDFG 1955b). Although the
14 San Joaquin River also supported a fall-run Chinook salmon run, they historically
15 composed a smaller portion of the river's salmon runs (Moyle 2002). Fall-run Chinook
16 salmon historically spawned in the main stem San Joaquin River upstream from the
17 Merced River confluence and in the main stem channels of the major tributaries
18 (Yoshiyama et al. 1996). Currently, however, they are limited to the Merced, Stanislaus,
19 and Tuolumne rivers where they spawn and rear downstream from main stem dams
20 (SJRRP 2011b). CDFW has operated a barrier (Hills Ferry Barrier) during the fall-run
21 Chinook salmon spawning season (October to December) at the confluence of the
22 Merced River with the San Joaquin River since the early 1990s to prevent adult fall-run
23 Chinook salmon from migrating further up the San Joaquin River, including the Reach
24 4B/ESB Project area, into warmer temperatures and impassable barriers that prevent them
25 from accessing suitable spawning habitat in reach 1. However, the Hills Ferry Barrier has
26 been demonstrated to be an ineffective barrier that many adult fall-run Chinook Salmon
27 are able to migrate past (SJRRP 2012, SJRRP 2013).

28 Fall-run Chinook salmon currently is the most abundant and widespread salmon run in
29 California and is supported by five hatcheries releasing a combined total of
30 approximately 35 million juveniles each year (Mills et al. 1997, Huber and Carlson
31 2015). NMFS determined that listing this ESU as threatened was not warranted (64
32 Federal Regulation [FR] 50394–50415, September 16, 1999), but subsequently classified
33 it as a species of concern because of specific risk factors (69 FR 19975, April 15, 2004).
34 In 2008, a collapse of Central Valley fall-run Chinook salmon occurred that has been
35 attributed to several causes, including poor ocean conditions for rearing, freshwater water
36 withdrawals, negative hatchery effects, and ongoing degradation of freshwater and
37 estuarine habitats (Lindley et al. 2009).

38 Fall-run Chinook salmon exhibit similar life history strategies as spring-run (see spring-
39 run above), with a few differences. Fall-run Chinook salmon do not have a summer
40 holding period; instead, they migrate upstream during the fall and typically spawn from
41 October through December, peaking in early to mid-November in the San Joaquin River
42 tributaries (SJRRP 2011b). Unlike spring-run Chinook salmon, only a small percent of
43 fall-run exhibits a yearling life history strategy, and the majority emigrate as fry or smolts

1 during the winter or spring of the year they were born. Fall-run Chinook salmon fry
2 typically disperse downstream from January through March, whereas smolts primarily
3 migrate between March and June in the Central Valley (Brandes and McLain 2001).

4 Like spring-run Chinook salmon, fall-run are believed to have historically used the Reach
5 4B/ESB Project area as an adult upstream migration corridor and as a juvenile rearing
6 and migration corridor during downstream emigration (see spring-run above).

7 **California Central Valley Steelhead** Historical rainbow trout/steelhead
8 (*Oncorhynchus mykiss*) distribution in the upper San Joaquin River is unknown; however,
9 in rivers where they still occur, their distribution is skewed further upstream compared to
10 Chinook salmon (Voight and Gale 1998, as cited in McEwan 2001, Yoshiyama et al.
11 1996) and are typically tributary spawners (SJRRP 2011b). Lindley et al. (2006)
12 predicted the historical distribution of steelhead (the anadromous form of *O. mykiss*),
13 using an Intrinsic Potential habitat model. They found that at least 81 independent
14 populations of *O. mykiss* were widely distributed throughout the Central Valley, but
15 populations were relatively less abundant in San Joaquin River tributaries than in
16 Sacramento River tributaries because of natural barriers to migration. Additionally, many
17 small tributaries to the major San Joaquin River tributaries have too high a gradient or too
18 little flow to have supported steelhead; consequently, they likely were restricted to the
19 main stems and larger tributaries (Lindley et al. 2006). Around 80 percent of the
20 historical spawning and rearing habitat is now behind impassable dams, and 38 percent of
21 the populations identified by the model have lost their entire habitat (Lindley et al. 2006).

22 *Oncorhynchus mykiss* has two classifications: steelhead, which refers to the anadromous
23 form, and rainbow trout, which refers to the non-anadromous form. The anadromous
24 distinct population segment of *O. mykiss* was listed under the Federal ESA by NMFS (63
25 FR 13347, March 19, 1998 and 71 FR 834, January 5, 2006). The California Central
26 Valley steelhead DPS includes all naturally spawned populations of anadromous
27 steelhead below natural and human-made impassable barriers in the Sacramento and San
28 Joaquin rivers and their tributaries, excluding steelhead from San Francisco and San
29 Pablo bays and their tributaries. NMFS has concluded that populations of naturally
30 reproducing steelhead have been experiencing a long-term decline in abundance
31 throughout their range (SJRRP 2011a). Populations in the southern portion of the range
32 have experienced the most severe declines, particularly in streams from the Central
33 Valley south, where many stocks have been extirpated (NMFS 2014). Since the early
34 20th century, 23 naturally reproducing populations of steelhead are believed to have been
35 extirpated in the western United States. Many more are thought to be in decline in
36 Washington, Oregon, Idaho, and California. The decline of stocks in California has been
37 particularly steep. The only limited data available on steelhead numbers in the San
38 Joaquin River Basin come from CDFW kodiak trawling samples collected on the lower
39 San Joaquin River at Mossdale. These data suggest that steelhead numbers declined in the
40 early 1990s and remained low through 2002 (NMFS 2009).

41 In the Central Valley, adult steelhead migrate upstream beginning in June, peaking in
42 September, and continuing through February or March (Hallock et al. 1961, Bailey 1954,
43 McEwan and Jackson 1996). Spawning occurs primarily from January through March but

1 may begin as early as late December and may extend through April (Hallock et al. 1961,
2 as cited in McEwan and Jackson 1996). Although most steelhead die after spawning,
3 some adults are capable of returning to the ocean and migrating back upstream to spawn
4 in subsequent years. Eggs hatch after 20 to 100 days, depending on water temperature
5 (Shapovalov and Taft 1954).

6 Steelhead rear in freshwater before outmigrating to the ocean as smolts. The length of
7 time juveniles spend in freshwater appears to be related to growth rate (Peven et al.
8 1994). In warmer areas, where feeding and growth are possible throughout the winter,
9 steelhead may require a shorter period in freshwater before smolting (Sogard et al. 2012).
10 Juveniles typically remain in their natal streams for at least one summer, dispersing from
11 fry schools to establish feeding territories (Sogard et al. 2012). Peak feeding and
12 freshwater growth rates occur in late spring and early summer (Sogard et al. 2012).
13 Juveniles either overwinter in their natal streams, if adequate cover exists or disperse to
14 other streams as presmolts to seek more suitable winter habitat (Bjornn 1971; Dambacher
15 1991). When stream temperatures fall below about 45 °F (7.2°C) in the late fall to early
16 winter, steelhead enter a period of winter inactivity spent, hiding in the substrate or
17 closely associated with instream cover, during which time growth ceases (Everest and
18 Chapman 1972). Juveniles' winter hiding behavior reduces their metabolism and food
19 requirements and reduces their exposure to predation and high flows (Bustard and Narver
20 1975), but substantial mortality still appears to occur in winter.

21 Steelhead migrate downstream to the ocean as smolts, typically at a length of 5.85 to
22 7.80 inches (14.86 to 19.81 cm) (Meehan and Bjornn 1991). A length of 5.46 inches
23 (13.87 cm) is typically cited as the minimum size for smolting (Wagner et al. 1963;
24 Peven et al. 1994). Emigration appears to be more closely associated with size than with
25 age; 6 to 8 inches (15.24 to 20.32 cm) is the most common size of downstream migrants.
26 Downstream migration in unregulated streams has been correlated with spring freshets
27 (Reynolds et al. 1993). Most steelhead spend 1 to 3 years in the ocean, with smaller
28 smolts tending to remain in saltwater for a longer period than larger smolts (Chapman
29 1958). Larger smolts have been observed to experience higher ocean survival rates (Ward
30 and Slaney 1988, Bond et al. 2008).

31 Historically, steelhead may have utilized the Reach 4B/ESB Project area for juvenile
32 migration and rearing and as an adult migration corridor on their way to spawning
33 grounds in the upper reaches of the San Joaquin River. Similar to Chinook salmon, the
34 extensive slough and off-channel aquatic habitat present historically in the Reach
35 4B/ESB Project area (see Historical Habitat section) likely provided excellent steelhead
36 rearing habitat (Jeffres et al. 2008). In the Sacramento River system, drifting
37 invertebrates, the primary prey of juvenile salmonids, have been found to be more
38 abundant on an inundated floodplain than in the adjacent river channel (Sommer et al.
39 2001).

40 **White Sturgeon** White sturgeon (*Acipenser transmontanus*) have a marine distribution
41 spanning from the Gulf of Alaska south to Mexico but a spawning distribution ranging
42 only from the Sacramento River northward (McCabe and Tracy 1994). Currently, self-
43 sustaining spawning populations are only known to occur in the Sacramento, Fraser, and

1 Columbia rivers. Landlocked populations are located above major dams in the Columbia
2 River basin, and residual non-reproducing fish above Shasta Dam and Friant Dam
3 occasionally have been found (SJRRP 2010a). In California, primary abundance is in the
4 San Francisco Estuary, with spawning occurring mainly in the Sacramento and Feather
5 rivers (Klimley et al. 2015). However, CDFG fisheries catch information obtained from
6 fishery report cards (CDFG 2008, 2009) documented 25 mature white sturgeon
7 encountered by fisherman in 2007 in the San Joaquin River, and 6 mature white sturgeon
8 encountered in 2008 upstream from Highway 140 (Reach 5). In addition, an unknown
9 number of white sturgeon were captured in the Restoration Area in 2009 (CDFG 2010).
10 In 2012, an adult white sturgeon was observed in Reach 5 with a dual frequency
11 identification sonar (SJRRP unpublished data). Adult sturgeon were caught in the sport
12 fishery industry in the San Joaquin River between Mossdale and the confluence with the
13 Merced River in late winter and early spring, suggesting this was a spawning run
14 (Kohlhorst 1976). Kohlhorst et al. (1991) estimated that approximately 10 percent of
15 the Sacramento River system spawning population migrated up the San Joaquin River.
16 According to Gruber et al (2012), white sturgeon were documented spawning in the
17 San Joaquin River just downstream of Laird Park at river kilometer (RK) 142 in April
18 2011, suggesting the San Joaquin River may be an important source of production for the
19 white sturgeon population in the Sacramento-San Joaquin river system. White sturgeon
20 were also documented spawning within a 24-kilometer reach of the San Joaquin River
21 from Sturgeon Bend (RK 119) to Grayson Road Bridge (RK 143) between March 20 and
22 May 14, 2012 (Jackson and Van Eenennaam 2013). Genetic analysis of wild white
23 sturgeon embryos collected during the 2012 spawning survey suggested that
24 approximately 40 individuals contributed to the 2012 spawning events out of less than
25 100 adults likely present in the San Joaquin River (Blankenship et al. 2017). However, in
26 subsequent San Joaquin River white sturgeon spawning surveys in the critical dry years
27 2013, 2014, and 2015, no white sturgeon eggs or larvae were captured despite the
28 presence of mature white sturgeon in the San Joaquin River (Heironomus et al. 2016,
29 Heironomus and Jackson 2017). The apparent negligible recruitment during critical dry
30 years is likely a result of poor water quality conditions in the San Joaquin River in critical
31 dry years, particularly low flows and high water temperatures during the spring spawning
32 period (Heironomus et al. 2016, Heironomus and Jackson 2017). In 2015, at least two of
33 the captured female sturgeon were undergoing atresia, the degeneration and resorption of
34 eggs, likely as a result of mean water temperatures remaining over 18°C for a week prior
35 to their capture (Heironomus and Jackson 2017). The spawning observations in wet 2011
36 and dry 2012 confirm that white sturgeon do spawn in the San Joaquin River in both wet-
37 and dry-year conditions (Jackson et al. 2016). In dry years, small magnitude, short
38 duration streamflow increases resulting from precipitation events or tributary river flow
39 pulses for juvenile salmonids appear to initiate white sturgeon spawning in the San
40 Joaquin River (Jackson et al. 2016).

41 White sturgeon spend most of their lives in estuaries of large rivers, only moving into
42 freshwater to spawn (Moyle 2002). Sturgeon migrate upstream when they are ready to
43 spawn in response to flow increases (Moyle 2002, Jackson et al. 2016). Male white
44 sturgeon are at least 10 to 12 years old before sexual maturity (Moyle 2002). Spawning
45 takes place between late February and early June when water temperatures range from 8
46 to 19°C (Moyle 2002). Telemetry studies in the San Joaquin River suggest a white

1 sturgeon spring migration and spawning from February through May (Heironimus and
2 Jackson 2017). The telemetry studies also suggest some fidelity to the San Joaquin River
3 with 37% (16 out of 43) of previously tagged fish (2012 to 2014) returning to the San
4 Joaquin River in 2015 (Heironimus and Jackson 2017). Large white sturgeon year classes
5 are associated with high outflows through the estuary in spring, presumably due to larval
6 sturgeon being moved quickly downstream to suitable rearing areas in the estuary (Moyle
7 2002).

8 Historically, white sturgeon likely only used the Reach 4B/ESB Project area as a
9 migration corridor during upstream spawning runs and downstream juvenile emigration.
10 Currently, numerous barriers in the Reach 4B/ESB Project area (see Stressors section)
11 likely act as complete barriers to adult sturgeon in most years.

12 **River Lamprey** River lampreys have been collected from large coastal streams from
13 20 kilometers north of Juneau, Alaska, to San Francisco Bay (Moyle 2002). In California,
14 most records are for the lower Sacramento-San Joaquin River system, including the
15 Stanislaus and Tuolumne rivers. The biology of river lamprey has not been well studied
16 in California with little primary literature available, so information available is based on
17 studies from British Columbia. Adults migrate into freshwater during the fall and spawn
18 during February through May in tributary streams. They dig saucer-shaped depressions in
19 gravelly riffles for spawning. Juvenile ammocoetes remain in silty backwaters and eddies
20 to feed on algae and microorganisms.

21 Due to the marshy, low gradient habitat present historically in the Reach 4B/ESB Project
22 area (see Historical Habitat section), river lamprey likely used this Reach for juvenile
23 rearing. However, due to several fish migration barriers present in the Reach 4B/ESB
24 Project area (see Stressors section), river lamprey likely are blocked from migrating
25 through the Reach 4B/ESB Project area or in reaches upstream in most years.

26 **Pacific Lamprey** Pacific lamprey (*Entosphenus tridentata*) are anadromous fish that
27 have Pacific coast distributions and have been found in the San Joaquin River (SJRRP
28 2017, SJRRP unpublished data - DNA barcoding analysis of lamprey). Pacific lamprey
29 does not appear to home to natal streams, as little genetic variation has been observed in
30 populations from British Columbia to southern California (Goodman et al. 2008). Instead,
31 they appear to key in on pheromones released by ammocoetes present in the river such
32 that they will not return to a river that lacks ammocoetes (Goodman and Reid 2012). The
33 result is a source-sink dynamic for Pacific lamprey such that large river systems
34 containing robust populations serve as sources for smaller rivers and streams that can be
35 sinks (Moyle et al. 2015). The Pacific lamprey has diverse life histories with some rivers
36 containing two runs; one run that returns in the spring and spawns immediately after
37 upstream migration and another run that migrates upstream in the fall and will spawn the
38 following spring (Moyle et al. 2015). Most adult Pacific lamprey spawning migrations
39 occur between March and late June, with upstream movement typically occurring during
40 the night (Moyle et al. 2015). Upstream migration seems to take place largely in response
41 to high flows, and adults can move substantial distances unless blocked by major barriers.
42 Due to several fish migration barriers present in the Reach 4B/ESB Project area (see
43 Stressors section), Pacific lamprey likely are blocked from migrating in the Reach

1 4B/ESB Project area or in reaches upstream in most years. However, Pacific lamprey
2 juveniles were found in Reach 1 during the 2013-2014 sampling demonstrating that
3 Pacific lamprey are able to migrate to Reach 1 in some years (SJRRP 2017).

4 Pacific lamprey hatching occurs in approximately 17 days at 57°F (14°C) and, after
5 spending an approximately equal period in redd gravels (Meeuwig et al. 2005),
6 ammocoetes (larvae) emerge and drift downstream to depositional areas where they
7 burrow into fine substrates and filter feed on organic materials (Moore and Mallatt 1980).
8 Throughout the ammocoete life stage, individuals will leave their burrows and drift to a
9 new area at night (Moyle et al. 2015). Ammocoetes remain in freshwater for 4 to 7 years
10 before undergoing a metamorphosis into an eyed, smolt-like form (macrophthalmia)
11 (Moore and Mallatt 1980, Moyle 2002, Moyle et al. 2015). At this time, individuals
12 migrate to the ocean between fall and spring, typically during winter and spring high-
13 flow events (Goodman et al. 2015), to feed parasitically on a variety of marine fishes and
14 smooth skinned marine mammals (Van de Wetering 1998, Moyle 2002). Pacific lamprey
15 remain in the ocean for approximately 18 to 40 months before returning to freshwater as
16 immature adults (Kan 1975, Beamish 1980). Pacific lampreys die soon after spawning,
17 though there is some anecdotal evidence that this is not always the case (Moyle 2002,).

18 Due to the marshy, low gradient habitat present historically in the Reach 4B/ESB Project
19 area (see Historical Habitat section), Pacific lamprey likely used this reach for juvenile
20 rearing. However, due to several Reach 4B/ESB Project area fish migration barriers (see
21 Stressors section), Pacific lamprey likely are blocked from migrating through the Reach
22 4B/ESB Project area or in reaches upstream in some to most years. However, juvenile
23 Pacific lamprey have been observed in Reach 1 (SJRRP 2017) demonstrating that adult
24 Pacific lamprey are able to pass all of the barriers in some years and would have had to
25 pass through reach 4B.

26 ***Native Riverine Fish Species***

27 Many of the native riverine species historically present in the Reach 4B/ESB Project area
28 are still present (CDFG 2007; SJRRP2017 Fish Assemblage Monitoring, Unpublished
29 Data), but their abundance trends are unknown. Historically, the San Joaquin River in
30 Reach 4B would have contained had year-round presence of the deep-bodied fishes
31 assemblage in addition to the anadromous salmonids which were migrating through
32 (Moyle 2002). The deep-bodied fish assemblage includes the Sacramento hitch,
33 Sacramento blackfish, Sacramento splittail, and Sacramento perch (Moyle 2002). Some
34 of these species still occur in Reach 4B while others do not (Table G-1). Degradation or
35 complete destruction of historical aquatic habitats due to dewatering, agricultural
36 conversion, levee construction, and channelization (see Stressors section), likely has led
37 to greatly reduced abundances of native riverine species in the Reach 4B/ESB Project
38 area. Furthermore, remaining native riverine species are likely competing with introduced
39 species for limited habitat (see Introduced Fish Species section).

40 **Sacramento Hitch** Sacramento Hitch are endemic to the Sacramento-San Joaquin River
41 Basin (SJRRP 2011b). There are three subspecies within this species found in the Clear
42 Lake, Pajaro, and Salinas watersheds and Sacramento-San Joaquin Watershed (Lee et al.
43 1980). Hitch occupy warm, low-elevation lakes, sloughs, and slow-moving stretches of

1 rivers and clear, low-gradient streams. Among native fishes, hitch have the highest
2 temperature tolerances in the Central Valley. They can withstand water temperatures up
3 to 100°F (38°C) although they prefer temperatures of 81 to 84°F (27 to 29°C). Hitch also
4 have moderate salinity tolerances and can be found in environments with salinities up to
5 9 parts per thousand (ppt) (Moyle 2002). Hitch require clean, smaller gravel and
6 temperatures of 57 to 64°F (14 to 18°C) to spawn. When larvae and small juveniles move
7 into shallow areas to shoal, they require vegetative refugia to avoid predators. Larger fish
8 are often found in deep pools containing an abundance of aquatic and terrestrial cover
9 (Moyle 2002).

10 Mass spawning migrations typically occur when flows increase during spring, raising
11 water levels in rivers, sloughs, ponds, reservoirs, watershed ditches, and riffles of lake
12 tributaries. Females lay eggs that sink into gravel interstices (SJRRP 2011b). Hatching
13 occurs in 3 to 7 days at 59 to 72°F (15 to 22°C), and larvae take another 3 to 4 days to
14 emerge. As they grow, they move into perennial water bodies where they would shoal for
15 several months in association with aquatic vegetation or other complex vegetation before
16 moving into open water. Hitch are omnivorous and feed in open waters on filamentous
17 algae, aquatic and terrestrial insects, zooplankton, aquatic insect pupae and larvae, and
18 small planktonic crustaceans (Moyle 2002).

19 **Sacramento blackfish** Sacramento blackfish are endemic to low-elevation portions of
20 major tributaries of the Sacramento and San Joaquin rivers (SJRRP 2011b). Although
21 they were abundant in the sizeable lakes of the historical San Joaquin Valley, they are
22 currently common only in sloughs and oxbow lakes of the Delta. Sacramento blackfish
23 are most abundant in warm, turbid, and often highly modified habitats.

24 They are found in locations ranging from deep turbid pools with clay bottoms to warm,
25 shallow, and seasonally highly alkaline water bodies. Blackfish have a remarkable ability
26 to adapt to extreme environments such as high temperatures and low dissolved oxygen
27 (DO) (Cech et al 1979, Campagna and Cech 1981). Although optimal temperatures range
28 from 72 to 82°F (22 to 28°C), adults frequently can be found in waters exceeding 86°F
29 (30°C). Their ability to tolerate extreme conditions affords them survival during periods
30 of drought or low flows (Moyle 2002).

31 Spawning occurs in shallow areas with dense aquatic vegetation between May and July
32 when water temperatures range between 54 and 75°F (12 to 24°C). Eggs attach to
33 substrate in aquatic vegetation, and larvae are frequently found in similar shallow areas.
34 Juvenile blackfish are often found in large schools within shallow areas associated with
35 cover and feed on planktonic algae and zooplankton (Moyle 2002).

36 **Sacramento splittail** Sacramento splittail are endemic to the Sacramento and San
37 Joaquin rivers, Delta, and San Francisco Bay (SJRRP 2011b). In the San Joaquin River,
38 they have been documented as far upstream as the town of Friant (Rutter 1908). In recent
39 wet years, splittail have been found as far upstream as Salt Slough (Saiki 1984, Baxter
40 2000) where the presence of both adults and juveniles indicated successful spawning.

1 Adult splittail move upstream in late November through late January, foraging in flooded
2 areas along the main rivers, bypasses, and tidal freshwater marsh areas before spawning
3 (Moyle et al. 2004). Feeding in flooded riparian areas before spawning may contribute to
4 spawning success and survival of adults after spawning (Moyle et al. 2004). Splittail
5 appear to concentrate their reproductive effort in wet years when potential success is
6 greatly enhanced by the availability of inundated floodplain habitat (Meng and Moyle
7 1995, Sommer et al. 1997). Splittail are fractional spawners, with individuals spawning
8 over several months (Wang 1995).

9 Eggs begin to hatch in 3 to 7 days, depending on temperature (Bailey 1994). After
10 hatching, the swim bladder inflates and larvae begin active swimming and feeding
11 (Moyle 2002). Most larval splittail remain in flooded riparian areas for 10 to 14 days,
12 most likely feeding in submerged vegetation before moving into deeper water as they
13 become stronger swimmers (Wang 1986, Sommer et al. 1997). Most juveniles move
14 downstream in response to flow pulses into shallow, productive bay and estuarine waters
15 from April to August (Meng and Moyle 1995, Moyle 2002). Floodplain habitat offers
16 high-quality food and production and low predator densities to increase juvenile growth
17 and survival.

18 Non-breeding splittail are found in temperatures up to 75°F (24°C) (Young and Cech
19 1996). Juveniles and adults have optimal growth at 68°F (20°C), with physiological
20 distress above 84°F (29°C) (Young and Cech 1996). Splittail have a high tolerance for
21 variable environmental conditions (Young and Cech 1996, Moyle et al. 2015) and are
22 generally opportunistic feeders. Prey includes mysid shrimp, clams, and some terrestrial
23 invertebrates.

24 **Hardhead** Hardhead are endemic to larger low- and mid-elevation streams of the
25 Sacramento-San Joaquin river basins (SJRRP 2011b). Hardhead are widely distributed in
26 foothill streams and may be found in a few reservoirs on the San Joaquin River upstream
27 from Millerton Lake. Hardhead prefer water temperatures above 68°F (20°C), with
28 optimal temperatures between 75 and 82°F (24 to 28°C). Their distribution is limited to
29 well-oxygenated streams and the surface water of impoundments. They are often found in
30 clear, deep pools greater than 31.5 inches (800 mm) and runs with slower water
31 velocities. Larvae and post-larvae may occupy river edges or flooded habitat before
32 seeking deeper low-velocity habitat as they increase in size (Moyle 2002).

33 Hardhead spawn between April and August. Females lay eggs on gravel in riffles, runs,
34 or the heads of pools. The early life history of hardhead is not well known. Juveniles may
35 feed on insects from the surface, whereas adults are benthivores, occupying deep pools.
36 Prey items may include insect larvae, snails, algae, aquatic plants, crayfish, and other
37 large invertebrates (Moyle 2002).

38 **Sacramento pikeminnow** Sacramento pikeminnow are endemic to the Sacramento-San
39 Joaquin River Basin (Moyle 2002). Sacramento pikeminnow prefer rivers in low- to mid-
40 elevation areas with clear water, deep pools, low-velocity runs, undercut banks, and
41 vegetation. They are not typically found where centrarchids have become established.

1 Sacramento pikeminnow prefer summer water temperatures above 59°F (15°C), with a
2 maximum of 79°F (26°C) (Moyle 2002).

3 Sexually mature fish move upstream in April and May when water temperatures are 59 to
4 68°F (15 to 20°C). Sacramento pikeminnow spawn over riffles or the base of pools in
5 smaller tributaries. Pikeminnow are slow growing and may live longer than 12 years.
6 Before the introduction of larger predatory fishes, pikeminnows may have been the apex
7 predator in the Central Valley. Pikeminnow prey includes insects, crayfish, larval and
8 mature fish, amphibians, lamprey ammocoetes, and occasionally small rodents (Moyle
9 2002).

10 **Sacramento sucker** Sacramento suckers have a wide distribution in California,
11 including streams and reservoirs of the Sacramento and San Joaquin watersheds (Moyle
12 2002). Sacramento suckers most commonly are found in cold, clear streams and
13 moderate-elevation lakes and reservoirs. Sacramento suckers can make relatively large
14 migrations related to spawning and flow variability (Jeffres et al. 2006). Shifts in
15 microhabitat use occur with smaller fish using shallow, low-velocity peripheral zones
16 moving to areas of deeper water as they grow (Cech et al. 1990). Sacramento suckers can
17 tolerate a wide range of temperature fluctuations, from streams that rarely exceed 59°F
18 (15°C) to those that reach up to 86°F (30°C). They have high salinity tolerances, having
19 been found in reaches with salinities greater than 13 ppt. Sacramento suckers can
20 colonize new habitats readily (Moyle 2002).

21 Sacramento suckers typically feed nocturnally on algae, detritus, and small benthic
22 invertebrates. They spawn over riffles from February through June when temperatures are
23 approximately 54 to 64°F (12 to 18°C). After embryos hatch in 2 to 4 weeks, larvae
24 remain close to the substrate until they are swept into warm, shallow water or among
25 flooded vegetation (Moyle 2002).

26 **Tule perch** Endemic Sacramento-San Joaquin River subspecies of tule perch
27 historically were widespread throughout the lowland rivers and creeks in the Central
28 Valley (SJRRP 2011b). Currently, in the San Joaquin River watershed, they occur in the
29 Stanislaus River, occasionally in the San Joaquin River near the Delta, and the lower
30 Tuolumne River. Tule perch in riverine habitat usually are found in emergent plant beds,
31 deep pools, and near banks with complex cover. They require cool, well-oxygenated
32 water, and tend not to be found in water exceeding 77°F (25°C) for extended periods.
33 They can tolerate high salinities (i.e., 30 ppt) (Moyle 2002).

34 Tule perch generally feed on the bottom or among aquatic plants (Moyle 2002). They are
35 primarily adapted to feed on small invertebrates and zooplankton. Females mate multiple
36 times between July and September, and sperm is stored until January when internal
37 fertilization occurs. Young develop within the female and are born in June or July when
38 food is most abundant. Juveniles begin to school soon after birth.

39 **Prickly sculpin** Central Valley populations of prickly sculpin (*Cottus asper*) are found
40 in the San Joaquin Valley south to the Kings River (Moyle 2002). Prickly sculpin
41 generally is found in medium-sized, low-elevation streams with clear water and bottoms

1 of mixed substrate and dispersed woody debris. In the San Joaquin Valley, they are
2 absent from warm, polluted areas, implying their distribution is regulated by water
3 quality. Prickly sculpin has been found in abundance in cool flowing water near Friant
4 Dam, in Millerton Lake, and in the small, shallow Lost Lake where bottom temperatures
5 exceed 79°F (26°C) in the summer (Moyle 2002).

6 Prickly sculpin spawn from February through June when water temperatures reach 46 to
7 55°F (8 to 13°C). After hatching, larvae move down into large pools, lakes, and estuaries
8 where they spend 3 to 5 weeks as planktonic fry. Prickly Sculpin prey include large
9 benthic invertebrates, aquatic insects, molluscs, and small fish and frogs (Moyle 2002).

10 **Riffle sculpin** Riffle sculpin (*Cottus gulosus*) have a scattered distribution pattern
11 throughout California, including the Sacramento-San Joaquin watersheds (Moyle 2002).
12 Riffle sculpin prefer habitats that are fairly shallow with moderately swift water
13 velocities and oxygen levels near saturation (Moyle and Baltz 1985). They move where
14 water temperatures do not surpass 77 to 79°F (25 to 26°C) and temperatures greater than
15 86°F (30°C) are generally lethal (Moyle 2002).

16 Riffle sculpins are benthic, opportunistic feeders (Moyle 2002). Spawning occurs
17 between February and April, with eggs deposited on the underside of rocks in swift riffles
18 or inside cavities of submerged logs. Eggs hatch in 11 to 24 days, and when fry reach
19 approximately 0.25 inches (6 mm) total length, they become benthic (Moyle 2002).

20 **Threespine stickleback** Central Valley populations of threespine stickleback
21 (*Gasterosteus aculeatus*) are scattered from the Lower Kings River and the San Joaquin
22 River below Friant Dam to roughly Redding in the Sacramento River drainage (Moyle
23 2002). Threespine sticklebacks are quiet-water fish, living in shallow, weedy pools and
24 backwaters or among emergent plants at stream edges over bottoms of gravel, sand, and
25 mud. Threespine sticklebacks are capable of completing their entire life cycle in either
26 freshwater or saltwater, migrating between the two environments.

27 In some areas, pikeminnow predation largely eliminated sticklebacks (Moyle 2002). This
28 may explain in part the scattered distribution of sticklebacks in many California River
29 systems, including those of the Central Valley. For example, in San Francisco Bay
30 streams, they are largely absent from areas containing introduced predatory fish.

31 **Kern brook lamprey** Kern brook lamprey (*Lampetra hubbsi*) are endemic to the
32 eastern portion of the San Joaquin Valley and were first collected in the Friant-Kern
33 Canal. They subsequently have been found in the lower Merced, Kaweah, Kings, and San
34 Joaquin rivers. They are generally found in silty backwaters of rivers stemming from the
35 Sierra foothills. The nonpredatory, resident Kern brook lamprey has not been studied
36 extensively, but it presumably has a similar life history and habitat requirements to the
37 western brook lamprey (*Lampetra richardsoni*) and other brook lamprey species. Like
38 other lampreys, the Kern brook lamprey is thought to spawn in the spring and die soon
39 thereafter (Moyle 2002). After eggs hatch, they remain in gravel redds until their yolk
40 sacs are absorbed. At this time, larvae emerge and drift downstream into low-velocity,
41 depositional rearing areas where they feed by filtering organic matter from the substrate.

1 After reaching approximately 4 to 6 inches (102 to 152 mm), ammocoetes undergo
2 metamorphosis into eyed adults (Moyle 2002). As with other brook lamprey species,
3 adults do not eat and may even shrink following metamorphosis (Moyle et al. 2015).
4 Adults prefer riffles containing small gravel for spawning and cobble for cover (Moyle
5 2002).

6 ***Non-native Introduced/Invasive Anadromous Fish Species***

7 **Striped Bass** Striped bass were first introduced in the San Francisco Bay in 1879 and
8 are now widely distributed throughout the Sacramento-San Joaquin drainage as far
9 upstream as fish barrier dams (Moyle 2002). Striped bass move regularly between salt
10 and fresh water, and they usually spend much of their life cycle in estuaries. Striped bass
11 are gregarious pelagic predators, reflected in their streamlined body shape, silvery
12 coloration, and feeding habits. Larval and juvenile striped bass are primarily invertebrate
13 feeders. As adults, striped bass are largely opportunistic feeders, with almost any fish
14 inhabiting the same area appearing in their diet. Striped bass are documented predators of
15 juvenile Chinook Salmon in the lower San Joaquin River (Demetras et al. 2016).
16 Predators in the lower San Joaquin River, including striped bass, were associated with
17 pools that were greater than 5 m deep (Cutter et al. 2017). Adult striped bass often reside
18 near diversion dams and other manmade structures which concentrate and may disorient
19 prey fish including juvenile salmonids (Sabal et al. 2016).

20 ***Non-native Introduced/Invasive Riverine Fish Species***

21 **Catfish species** Several species of catfish have been introduced into the Reach 4B/ESB
22 Project area, including black bullhead (*Ameiurus melas*), brown bullhead (*Ameiurus*
23 *nebulosus*), channel (*Ictalurus punctatus*), and white catfish (*Ameiurus catus*). Catfish
24 prefer slow moving, warm water habitat, are opportunistic omnivores, and scavenge off
25 the bottom of their habitat (Moyle 2002). Juvenile catfish mainly feed on crustaceans and
26 the larvae of aquatic insects. As catfish grow larger, other fish and crayfish become
27 increasingly important food sources. Although their interaction with native fishes has not
28 been studied (Moyle 2002), they likely directly compete for resources with native
29 bottom-feeding species such as hardhead, California roach, and Sacramento Sucker. The
30 common predators of juvenile Chinook salmon in the lower San Joaquin River include
31 channel and white catfish (Cutter et al. 2017).

32 **Forage fish species** Several small, forage fish species have been introduced into the
33 Reach 4B/ESB Project area, including bigscale logperch (*Percina macrolepida*), fathead
34 minnow (*Pimephales promelas*), inland silverside (*Menidia beryllina*), red shiner
35 (*Cyprinella lutrensis*), shimofuri goby (*Tridentiger bifasciatus*), golden shiner
36 (*Notemigonus crysoleucas*), goldfish (*Carassius auratus*), and western mosquitofish
37 (*Gambusia affinis*). Introduced forage fish species likely compete most with native
38 Sacramento splittail and the larval and juvenile life stages of many native fish species that
39 rely on zooplankton for prey.

40 **Common Carp** In California, common carp (*Cyprinus carpio*) are present across the
41 Sacramento-San Joaquin drainage (Moyle 2002). Common carp are most abundant in

1 warm, turbid water where habitat with silty bottoms and growths of submergent and
 2 emergent vegetation dominate. Common carp can tolerate a wide range of turbidities,
 3 temperatures, oxygen concentrations, and salinities. In general, common carp are
 4 omnivorous bottom feeders, particularly favoring insect larvae and small mollusks. Carp
 5 typically root around on silty bottoms, stirring up aquatic insects, which they then pick
 6 from the water. Through this foraging behavior, they can decrease local water clarity and
 7 prevent dense beds of aquatic plants from growing.

8 **Bass and Sunfish species** Several species of bass and sunfish have been introduced into
 9 the Reach 4B/ESB Project area, including black crappie (*Pomoxis nigromaculatus*), white
 10 crappie (*Pomoxis annularis*), bluegill (*Lepomis macrochirus*), green sunfish (*Lepomis*
 11 *cyaneus*), pumpkinseed (*Lepomis gibbosus*), redear sunfish (*Lepomis microlophus*),
 12 warmouth (*Lepomis gulosus*), spotted bass (*Micropterus punctulatus*), and largemouth
 13 bass (*Micropterus salmoides*). Bass and sunfish species prefer lakes, ponds, or low-
 14 velocity habitat in rivers (Moyle 2002). In the lower San Joaquin River, largemouth bass
 15 appear to be associated with submerged aquatic vegetation in pools (Cutter et al. 2017).
 16 Sunfish prefer habitats with aquatic vegetation and spawn in a variety of substrates. They
 17 prefer water temperatures above 27°C. Juvenile bass tend to feed on invertebrates,
 18 whereas adults are predominantly piscivorous. Sunfish are opportunistic feeders and eat a
 19 variety of aquatic insects, fish eggs, and planktonic crustaceans. Bass and sunfish species
 20 likely prey upon the larval and juvenile life stages of many native fish species present in
 21 the Reach 4B/ESB Project area.

22 **Threadfin shad** Threadfin shad were first introduced into California waters in the
 23 1950s and have since become established in the Sacramento-San Joaquin drainage
 24 (Moyle 2002). Threadfin shad inhabit open waters of reservoirs, lakes, and large ponds as
 25 well as sluggish backwaters of rivers. Threadfin shad are planktonic feeders and use their
 26 gill rakers to strain small zooplankton, phytoplankton, and detritus particles from the
 27 water while also feeding individually on larger zooplankton organisms.

28 G.2 References

- 29 Araki, H., B.A. Berejikian, M.J. Ford, and M.S. Bouin. 2008. *Fitness of hatchery-reared*
 30 *salmonids in the wild*. Evolutionary Applications 1: 342-355.
- 31 Bader, J.A., S.A. Moore, and K.E. Nusbaum. 2006. *The effect of cutaneous injury on a*
 32 *reproducible immersion challenge for Flavobacterium columnare infection in*
 33 *channel catfish (Ictalurus punctatus)*. Aquaculture 253: 1-9.
- 34 Bailey, E.D. 1954. *Time pattern of 1953-54 migration of salmon and steelhead into the*
 35 *upper Sacramento River*. Unpublished report. California Department of Fish and
 36 Game.
- 37 Bailey, H.C. 1994 *Sacramento splittail work continues at UC Davis*. Interagency
 38 Ecological Program Newsletter 7 (3):3.

- 1 Bams, R.A. 1970. *Evaluation of a revised hatchery method tested on pink and chum*
2 *salmon fry*, Journal of the Fisheries Research Board of Canada: 27: 1429–1452.
- 3 Baxter, R.D. 2000. *Splittail and longfin smelt*. IEP Newsletter 13: 19–21.
- 4 Beamesderfer, R. C. P., and M. A. H. Webb. 2002. *Green sturgeon status review*
5 *information*. S. P. Cramer & Associates. Gresham, Oregon. Prepared for State
6 Water Contractors. April 26.
- 7 Beamish, R.J. 1980. *Adult biology of the River Lamprey (Lampetra ayresi) and the*
8 *Pacific lamprey (Lampetra tridentata) from the Pacific coast of Canada*.
9 Canadian Journal of Fisheries and Aquatic Science 37: 1906–1923.
- 10 Beckon, W. 2007. *Selenium risk to salmonids with particular reference to the Central*
11 *Valley of California*. Poster presented at the American Fisheries Society 137th
12 Annual Meeting, San Francisco, California. September 2-6, 2007. U.S. Fish and
13 Wildlife Service, Sacramento, California.
- 14 Beechie, T.J., M. Liermann, E.M. Beamer, and R. Henderson. 2005. *A classification of*
15 *habitat types in a large river and their use by juvenile salmonids*. Transactions of
16 the American Fisheries Society 134: 717-729.
- 17 Beechie T.J. and T.H. Sibley. 1997. *Relationships between channel characteristics,*
18 *woody debris, and fish habitat in northwestern Washington streams*. Transactions
19 of the American Fisheries Society 126: 217-229.
- 20 Bellmore, J. Ryan; Baxter, Colden V.; Martens, Kyle; and Connolly, Patrick J., 2013. *The*
21 *floodplain food web mosaic: a study of its importance to salmon and steelhead*
22 *with implications for their recovery*. USGS Staff -- Published Research: Paper
23 699.
- 24 Berejikian, B.A., S.B. Matthews, and T.P. Quinn. 1996. *Effects of hatchery and wild*
25 *ancestry and rearing environments on the development of agnostic behavior in*
26 *steelhead trout (Oncorhynchus mykiss) fry*. Canadian Journal of Fisheries and
27 Aquatic Sciences 53:2004-2014.
- 28 Bjornn, T. C. 1971. *Trout and salmon movements in two Idaho streams as related to*
29 *temperature, food, stream flow, cover, and population density*. Transactions of the
30 American Fisheries Society 100:423–438.
- 31 Bjornn, T.C., and D.W. Reiser. 1991. *Habitat requirements of salmonids in streams*.
32 *Pages 83–138 in W. R. Meehan, editor*. Influences of forest and rangeland
33 management on salmonid fishes and their habitats. American Fisheries Society
34 Special Publication No. 19.
- 35 Blankenship, S.M., G. Schumer, J.P. Van Eenennaam, and Z.J. Jackson. 2017. Estimating
36 number of spawning white sturgeon adults from embryo relatedness. Fisheries
37 Management and Ecology 24: 163-172.

- 1 Boles, G. 1988. *Water temperature effects on Chinook salmon (Oncorhynchus*
2 *tshawytscha) with emphasis on the Sacramento River: A literature review.*
3 California Department of Water Resources, Northern District. 42 pp.
- 4 Bond, M.H., S.A. Hayes, C.V. Hanson, and R.B. MacFarlane. 2008. Marine survival of
5 steelhead (*Oncorhynchus mykiss*) enhanced by a seasonally close estuary.
6 Canadian Journal of Fisheries and Aquatic Sciences 65: 2242-2252.
- 7 Brandes, P.L., and J.S. McLain. 2001. *Juvenile Chinook salmon abundance, distribution,*
8 *and survival in the Sacramento-San Joaquin Estuary.* Pages 39-138 in Brown,
9 R.L., editor. Fish Bulletin 179: Contributions to the biology of Central Valley
10 salmonids. Volume 2. California Department of Fish and Game, Sacramento,
11 California.
- 12 Brekke, L. D., Miller, N. L., Bashford, K. E., Quinn, N. W., & Dracup, J. A. 2004.
13 *Climate change impacts uncertainty for water resources in the San Joaquin River*
14 *Basin, California.* Journal of the American Water Resources Association, 40(1),
15 149-164.
- 16 Brown, L. 1997. *Concentrations of chlorinated organic compounds in biota and bed*
17 *sediment in streams of the San Joaquin Valley, California.* Archives of
18 Environmental Contamination and Toxicology 33: 357-368.
- 19 Brown, L., and J.T. May. 2000. *Macroinvertebrate assemblages on woody debris and*
20 *their relations with environmental variables in the lower Sacramento and San*
21 *Joaquin river drainages, California: Environmental Monitoring and Assessment*
22 64: 311-329.
- 23 Brown, L.R. 1998. *Assemblages of fishes and their associations with environmental*
24 *variables, lower San Joaquin River drainage, California.* Open-File Report 98-
25 77. U.S. Geological Survey, National Water-Quality Assessment Program,
26 Sacramento, California.
- 27 _____. 2000. *Fish communities and their associations with environmental variables,*
28 *lower San Joaquin River drainage, California.* Environmental Biology of Fishes
29 57:251-269.
- 30 Brown, L.R., C.R. Kratzer and N.M. Dubrovsky. 1999. *Integrating chemical, water*
31 *quality, habitat, and fish assemblage data from the San Joaquin River drainage,*
32 *California.* In: C. Smith and K. Scow (ed.) *Integrated Assessment of Ecosystem*
33 *Health, CRC Press, Boca Raton (in press).*
- 34 Bustard, D. R., and D. W. Narver. 1975. *Aspects of the winter ecology of juvenile coho*
35 *salmon (Oncorhynchus kisutch) and steelhead trout (Salmo gairdneri).* Journal of
36 the Fisheries Research Board of Canada 32:667-680.

37

- 1 California Department of Fish and Game (CDFG). 1955a. *DFG testimony for a DWR*
2 *hearing on San Joaquin River water applications*. The Salmon Fishery of the San
3 Joaquin River, California: its history, its destruction, and its possible re-
4 establishment. Term paper, David Cone, 1973.
- 5 _____. 1955b. Report on water right applications 23, 234, 1465, 5638, 5817, 5818, 5819,
6 5820, 5821, 5822, 9369, United States of America – Bureau of Reclamation;
7 water right applications 6771, 6772, 7134, 7135, City of Fresno; water right
8 application 6733 – Fresno Irrigation District on the San Joaquin River,
9 Fresno/Madera, and Merced counties, California, DFG, Region 4, Fresno,
10 California.
- 11 _____. 1998. *A status review of the spring-run Chinook salmon (Oncorhynchus*
12 *tshawytscha) in the Sacramento River drainage*. Candidate Species Report 98-01.
13 Sacramento, California.
- 14 _____. 2007. *San Joaquin River fishery and aquatic resources inventory*. Final report,
15 September 2003 – September 2005.
- 16 _____. 2008. Gleason, E., M. Gingras, and J. DuBois. 2007 *Sturgeon fishing report*
17 *card: preliminary data report*. Stockton, California: California Department of
18 Fish and Game, Bay Delta Region.
- 19 _____. 2009. DuBois, J., M. Gingras, and R. Mayfield. 2008 *Sturgeon fishing report*
20 *card: preliminary data report*. Stockton, California: California Department of
21 Fish and Game, Bay Delta Region. June 17.
- 22 _____. 2010. DuBois, J., T. Matt, and B. Beckett. 2009 *Sturgeon fishing report card:*
23 *preliminary data report*. Stockton, California: California Department of Fish and
24 Game. Bay Delta Region (East). Available at:
25 ftp://ftp.delta.dfg.ca.gov/Adult_Sturgeon_and_Striped. March 29.
- 26 California Department of Fish and Wildlife (CDFW). 2016. Instream flow evaluation of
27 upstream spring-run Chinook salmon passage in Butte Creek, California.
28 Department of Fish and Wildlife Stream Evaluation Report No. 16-1. 133 p.
- 29 Campagna C. G., J. J. Cech Jr. 1981. *Gill ventilation and respiratory efficiency of*
30 *Sacramento blackfish, Orthodon microlepidotus in hypoxic environments*. Journal
31 of Fish Biology 19:581–591.
- 32 Cech, J.J., Mitchell, S.J., Massingill, M.J., 1979. *Respiratory adaptations of Sacramento*
33 *blackfish, Orthodon microlepidotus (Ayres), for hypoxia. Comparative*
34 *Biochemistry and Physiology 63A, 411-415.*
- 35 Cech, J.J., Jr., S.J. Mitchell, D.T. Castleberry, and M. McEnroe. 1990. *Distribution of*
36 *California stream fishes: influence of environmental temperature and hypoxia.*
37 *Environmental Biology of Fishes 29:95-105.*

- 1 Central Valley Regional Water Quality Control Board (Central Valley RWQCB). 1998.
2 *The Water Quality Control Plan (basin plan) for the California Regional Water*
3 *Quality Control Board Central Valley Region, Fourth Edition (2016*
4 *Amendments), The Sacramento River Basin and the San Joaquin River Basin.*
5 *California Regional Water Quality Control Board Central Valley Region,*
6 *Sacramento, California.*
- 7 _____ . 2001. *San Joaquin River Selenium TMDL*. Central Valley Regional Water
8 Quality Control Board, Rancho Cordova, California.
9 <http://www.waterboards.ca.gov/centralvalley/programs/tmdl/selenium.htm>.
- 10 Chapman, D. W. 1958. *Studies on the life history of Alsea River Steelhead*. Journal of
11 Wildlife Management 22(2):123–134.
- 12 Clark, G.H. 1943. Salmon at Friant Dam - 1942. *California Fish and Game* 29(3):89-91.
- 13 Collins, B.D., D.R. Montgomery, and A.D. Haas. 2002. *Historical changes in the*
14 *distribution and functions of large wood in Puget Lowland rivers*. Canadian
15 Journal of Fisheries and Aquatic Sciences 59: 66-76.
- 16 Cramer Fish Sciences. 2006. *2005-06 Stanislaus River weir data report*. Final report
17 prepared for the Anadromous Fish Restoration Program. June.
- 18 _____ . 2007. *Upstream fish passage at a resistance board weir Using infrared and*
19 *digital technology in the lower Stanislaus River, California, 2006–2007 Annual*
20 *Data Report*. Report prepared by Jesse T. Anderson, Clark B. Watry, and Ayesha
21 Gray for the Anadromous Fish Restoration Program.
- 22 Cutter, G.R., S.C. Manugian, J. Renfree, J. Smith, M.D. Huff, T.S. Sessions, B.E. Elliot,
23 K. Stierhoff, S. Mau, D. Murfin, and D.A. Demer. 2017. Mobile acoustic
24 sampling to map bathymetry and quantify the densities and distributions of
25 salmonid smolt predators in the San Joaquin River. NOAA Technical
26 Memorandum, NOO-TM-NMFS-SWFSC-575. 134 pp.
- 27 Dambacher, J. M. 1991. *Distribution, abundance, and emigration of juvenile steelhead*
28 *(Oncorhynchus mykiss), and analysis of stream habitat in the Steamboat Creek*
29 *Basin, Oregon. Master's thesis*. Oregon State University. Corvallis, Oregon.
- 30 Demetras, N.J., D.D. Huff, C.J. Michel, J.M. Smith, G.R. Cutter, S.A. Hayes, and S.T.
31 Lindley. 2016. *Development of underwater recorders to quantify predation of*
32 *juvenile Chinook salmon (Oncorhynchus tshawytscha) in a river environment*.
33 Fishery Bulletin 114: 179-185.
- 34 Demko D.B., C. Gemperle, S.P. Cramer, and A. Phillips. 1998. *Evaluation of juvenile*
35 *Chinook behavior, migration rate and location of mortality in the Stanislaus River*
36 *through the use of radio tracking*. Report prepared for Tri-dam Project. Gresham,
37 Oregon. December 1998.

Reach 4B, Eastside Bypass, and Mariposa Bypass
Channel and Structural Improvements Project

- 1 Domagalski JL, Dubrovsky NM, Kratzer CR. 1997. *Pesticides in the San Joaquin River,*
2 *California: inputs from the dormant sprayed orchards.* Journal of Environmental
3 Quality, 26, 454–465.
- 4 Domagalski, J.L., D.L. Knifong, P.D. Dileanis, L.R. Brown, J.T. May, V. Connor, and
5 C.N. Alpers. 2000. *Water quality in the Sacramento River Basin, California,*
6 *1994-98.* Circular 1215. USGS, National Water Quality Assessment Program.
7 Available at <http://pubs.usgs.gov/circ/circ1215/>.
- 8 Dubrovsky, N.M., C.R. Kratzer, L.R. Brown, J.M. Gronberg, and K.R. Burow. 1998.
9 *Water quality in the San Joaquin-Tulare basins, California, 1992-95.* USGS
10 Circular 1159. U.S. Geological Survey, Denver, Colorado.
- 11 Evans, A. F., et al. 2011. *Recovery of coded wire tags at a Caspian Tern colony in San*
12 *Francisco Bay: a technique to evaluate impacts of avian predation on juvenile*
13 *salmonids.* North American Journal of Fisheries Management 31: 79-87
- 14 Evans, A. F., Hostetter, N. J., et al. 2012. *Systemwide evaluation of avian predation of*
15 *juvenile salmonids from the Colombia River based on recoveries of passive*
16 *integrated transponder tags.* Transactions of the American Fisheries Society 141:
17 975-989.
- 18 Everest, F.H., and D.W. Chapman. 1972. *Habitat selection and spatial interaction by*
19 *juvenile Chinook salmon and steelhead trout in two Idaho streams.* Journal of the
20 Fisheries Research Board of Canada 29: 91-100.
- 21 Fisher, F.W. 1994. *Past and present status of Central Valley Chinook salmon,*
22 *Conservation Biology 8: 870–873.*
- 23 Florsheim, J.L., J.F. Mount, and A. Chin. 2008. *Bank erosion as a desirable attribute of*
24 *rivers.* Bioscience 58: 519-529.
- 25 Foott, J.S. and A. Imrie. 2016. *Prevalence and severity of Ceratonova shasta and*
26 *Parvicapsula minibicornis infection of natural Feather River juvenile Chinook*
27 *Salmon (January to May 2016).* U.S. Fish and Wildlife Service California-Nevada
28 Fish Health Center, Anderson, CA.
- 29 Foott, J.S. and A. Imrie. 2017. *Prevalence and severity of Ceratonova shasta and*
30 *Parvicapsula minibicornis infection of natural Feather River juvenile Chinook*
31 *Salmon (January to May 2017).* U.S. Fish and Wildlife Service California-Nevada
32 Fish Health Center, Anderson, CA
- 33 Fry, D.H. Jr. 1961. *King salmon spawning stocks of the California Central Valley, 1940-*
34 *1959.* California Fish and Game 47: 55-71.
- 35 Gallagher, A.S. Barriers. In: Bain, M.B., Stevenson, N.J. (Eds). Aquatic Habitat
36 Assessment: Common Methods. American Fisheries Society, Bethesda, MD, pp.
37 135-147.

- 1 Gilliom, R.J. and D.G. Clifton. 1990. *Organochlorine pesticide residues in bed sediments*
2 *of the San Joaquin River, California*. Water Resources Bulletin 26:11-24.
- 3 Goodbred, S.L., R.J. Gilliom, T.S. Gross, N.P. Denslow, W.L. Bryant, and T.R. Schoeb.
4 1997. *Reconnaissance of 17 β -estradiol, 11-ketotestosterone, vitellogenin, and*
5 *gonad histopathology in common carp of United States streams: potential for*
6 *contaminant-induced endocrine disruption*. U.S. Geological Survey Open-File
7 Report 96-627. Sacramento, California.
- 8 Goodman, D.H. and S.B. Reid. 2012. *Pacific Lamprey (Entosphenus tridentatus)*
9 *assessment and template for conservation measures in California*. U.S. Fish and
10 Wildlife Service, Arcata, CA. 117 pp.
- 11 Goodman, D.H., S.B. Reid, M.F. Docker, G.R. Haas, and A.P. Kinziger. 2008.
12 *Mitochondrial DNA evidence for high levels of gene flow among populations of a*
13 *widely distributed anadromous lamprey Entosphenus tridentatus*
14 *(Petromyzontidae)*. Journal of Fish Biology 72:400-417.
- 15 Goodman, D.H., S.B. Reid, N.A. Som, and W. R. Poytress. 2015. *The punctuated*
16 *seaward migration of Pacific lamprey (Entosphenus tridentatus): environmental*
17 *cues and implications for streamflow management*. Canadian Journal of Fisheries
18 and Aquatic Sciences 72(12):1817-1828.
- 19 Grossman, G.D., 2016. *Predation on Fishes in the Sacramento-San Joaquin Delta:*
20 *Current Knowledge and Future Directions*. San Francisco Estuary and Watershed
21 Science, 14(2).
- 22 Gruber, J. J., Z. J. Jackson, and J. P. Van Eenennaam. 2012. 2011 *San Joaquin River*
23 *sturgeon spawning survey*. Stockton Fish and Wildlife Office, Anadromous Fish
24 Restoration Program, U. S. Fish and Wildlife Service, Stockton, California.
- 25 Gurnell, A.M., H. Piegay, F.J. Swanson, and S.V. Gregory. 2002. *Large wood and fluvial*
26 *processes*. Freshwater Biology 47: 601-619.
- 27 Hallock, R.J., W.F. Van Woert, and L. Shapovalov. 1961. *An evaluation of stocking*
28 *hatchery-reared steelhead rainbow trout (Salmo gairdnerii gairdnerii) in the*
29 *Sacramento River system*. Fish Bulletin 114. California Department of Fish and
30 Game.
- 31 Hanson, C.H. 2001. Are juvenile Chinook salmon entrained at unscreened diversions in
32 direct proportion to the volume of water diverted. Pages 331-342 in Contributions
33 to the Biology of Central Valley Salmonids. Fish Bulletin 179: Volume 2.
- 34 Healey, M.C. 1991. *The life history of Chinook salmon. Pages 311-393 in C. Groot and*
35 *L Margolis, editors. Pacific salmon life histories*. University of British Columbia
36 Press, Vancouver, Canada.

- 1 Heironimus, L.B., and Z.J. Jackson. 2017. 2015 San Joaquin River white sturgeon
2 telemetry study. Lodi Fish and Wildlife Office, Anadromous Fish Restoration
3 Program, U.S. Fish and Wildlife Service, Lodi, CA.
- 4 Heming, T.A. 1982. *Effects of temperature on utilization of yolk by Chinook salmon*
5 *(Oncorhynchus tshawytscha) eggs and alevins*. Canadian Journal of Fisheries and
6 Aquatic Sciences 39: 184–190.
- 7 Hendrickson, G. L., A. Carleton, and D. Manzer. 1989. *Geographic and seasonal*
8 *distribution of the infective stage of Ceratomyxa shasta (Myxozoa) in Northern*
9 *California*. Diseases of Aquatic Organisms 7:165-169.
- 10 Herren, J.R., and S.S. Kawaski. 2001. *Inventory of water diversions in four geographic*
11 *areas in California's Central Valley*. Pages 343-3552 in Brown, R.L., editor. *Fish*
12 *Bulletin 179: Contributions to the biology of Central Valley salmonids*. Volume 2.
13 California Department of Fish and Game, Sacramento, California.
- 14 Huber, E.R. and S.M. Carlson. 2015. *Temporal trends in hatchery releases of fall-run*
15 *Chinook salmon in California's Central Valley*. San Francisco Estuary and
16 Watershed Science 13(2).
- 17 Jackson, Z.J. and J.P. Van Eenennaam. 2013. 2012. *San Joaquin River sturgeon*
18 *spawning survey*. Stockton Fish and Wildlife Office, Anadromous Fish
19 Restoration Program, U.S. Fish and Wildlife Service, Lodi, California.
- 20 Jackson, Z.J., J.J. Gruber, and J.P. Van Eenennaam. 2016. *White Sturgeon Spawning in*
21 *the San Joaquin River, California, and Effects of Water Management*. Journal of
22 Fish and Wildlife Management 7: 171-180.
- 23 Jeffres, C.A., A.P. Klimley, J.E. Merz, and J. J. Cech. 2006. *Movement of Sacramento*
24 *sucker, Catostomus occidentalis, and hitch, Lavinia exilicauda, during a spring*
25 *release of water from Camanche Dam in the Mokelumne River, California."*
26 *Environmental Biology of Fishes* 75, no. 4 (2006): 365-373.
- 27 Jeffres, C.A., J.J. Opperman, P.B. Moyle. 2008. *Ephemeral floodplain habitats provide*
28 *best growth conditions for juvenile Chinook salmon in a California River*.
29 *Environmental Biology of Fishes* 83:449–458.
- 30 Jones and Stokes. 2001. *Technical Memorandum on the Potential Barriers to Migrating*
31 *Steelhead and Chinook Salmon on the San Joaquin River*. Sacramento, California.
32 Prepared for Friant Water Users Authority and Natural Resources Defense
33 Council.
- 34 Kan, T.T. 1975. *Systematics, variation, distribution, and biology of lampreys of the genus*
35 *Lampetra in Oregon*. Dissertation for the Doctor of Philosophy. Oregon State
36 University, Corvallis, Oregon. 194 pp.

- 1 Katz, J.V.E, Jeffres, C., Conrad, J.L., Sommer, T.R., Martinez, J., Brumbaugh, S., et al.
2 (2017). *Floodplain farm fields provide novel rearing habitat for Chinook salmon*.
3 PLoS ONE 12(6): e0177409.
- 4 Klimley, A.P., Chapman, E.D., Cech Jr, J.J., Cocherell, D.E., Fangue, N.A., Gingras, M.,
5 Jackson, Z., Miller, E.A., Mora, E.A., Poletto, J.B. and Schreier, A.M. 2015.
6 *Sturgeon in the Sacramento–San Joaquin Watershed: New Insights to Support*
7 *Conservation and Management*. San Francisco Estuary and Watershed Science,
8 13(4).
- 9 Kohlhorst, D.W. 1976. *Sturgeon spawning in the Sacramento River in 1973*, as
10 determined by distribution of larvae. California Fish and Game 62:32-40.
- 11 Kohlhorst, D.W., L.W. Botsford, J.S. Brennan, and G.M. Cailliet, 1991. Aspects of the
12 structure and dynamics of an exploited central California population of white
13 sturgeon (*Acipenser transmontanus*), in "Acipenser," P. Wouldiot, ed., Bordeaux,
14 France, pp. 277 – 293.
- 15 Kostow, K. 2009. *Factors that contribute to the ecological risks of salmon and steelhead*
16 *hatchery programs and some mitigation strategies*. Reviews in Fish Biology and
17 Fisheries 19: 9-31.
- 18 Kratzer, C.R., and J.L. Shelton. 1998. *Water quality assessment of the San Joaquin -*
19 *Tulare basins, California: analysis of available data on nutrients and suspended*
20 *sediment in surface water, 1972-1990*. Professional Paper 1587. U.S. Geological
21 Survey, National Water-Quality Assessment Program, Sacramento, California.
- 22 Kuivila, K.M. and C.G. Foe. 1995. *Dormant spray pesticides in the San Francisco*
23 *Estuary, California*. Pages 72-73 in The Wildlife Society second annual
24 conference (abstracts). The Wildlife Society, Bethesda, Maryland.
- 25 Laikre, L., M.K. Schwartz, R.S. Waples, and N. Ryman. 2010. *Compromising genetic*
26 *diversity in the wild: unmonitored large-scale release of plants and animals*.
27 Trends in Ecology and Evolution 25: 520-529.
- 28 Latterell, J.L. and R.J. Naiman. 2007. *Sources and dynamics of large logs in a temperate*
29 *floodplain river*. Ecological Applications 17: 1127-1141.
- 30 Lee, D.S., C.R. Gilbert, C.H. Hocutt, R.E. Jenkins, D.E. McAllister, and J.R. Stauffer,
31 editors. 1980. *Atlas of North American freshwater fishes*. North Carolina State
32 Museum of Natural History, Raleigh, NC.
- 33 Levin, P.S., R.W. Zabel, and J.G. Williams. 2001. *The road to extinction is paved with*
34 *good intentions: negative association of fish hatcheries with threatened salmon*.
35 Proceedings of the Royal Society B 268: 1153-1158.
- 36 Limm, M. P., and Marchetti, M. P. 2009. Juvenile Chinook salmon (*Oncorhynchus*
37 *tshawytscha*) growth in off-channel and main-channel habitats on the Sacramento

- 1 River, CA using otolith increment widths. *Environmental biology of fishes* 85:
2 141-151.
- 3 Lindley, S.T., R. Schick, B.P. May, J.J. Anderson, S. Greene, C. Hanson, A. Low, D.
4 McEwan, R.B. MacFarlane, C. Swanson, and J.G. Williams. 2004. *Population*
5 *structure of threatened and endangered Chinook salmon ESUs in California's*
6 *Central Valley Basin*. Technical Memorandum NOAA-TM-NMFS-SWFSC-360.
7 National Marine Fisheries Service, Southwest Fisheries Science Center, Santa
8 Cruz, California.
- 9 Lindley, S.T., R.S. Schick, A. Agrawal, M. Goslin, T.E. Pearson, E. Mora, J.J. Anderson,
10 B. May, S. Greene, C. Hanson, A. Low, D. McEwan, R.B. MacFarlane, C.
11 Swanson, and J.G. WOULDiams. 2006. *Historical population structure of Central*
12 *Valley steelhead and its alteration by dams*. *San Francisco Estuary & Watershed*
13 *Science* 4(1):1–19.
- 14 Lindley, S. T., C.B. Grimes, M.S. Mohr, W. Peterson, J. Stein, J.T. Anderson, L.W.
15 Botsford, D.L. Bottom, C.A. Busack, T.K. Collier, J. Ferguson, J.C. Garza, A.M.
16 Grover, D.G. Hankin, R.G. Kope, P.W. Lawson, A. Low, R.B. MacFarlane, K.
17 Moore, M. Palmer-Zwahlen, F.B. Schwing, J. Smith, C. Tracy, R. Webb, B.K.
18 Wells, and T.H. Williams. 2009. *What caused the Sacramento River fall Chinook*
19 *stock collapse?* Technical Memorandum NOAA-TM-NMFS-SWFSC-447.
20 National Marine Fisheries Service, Southwest Fisheries Science Center, Santa
21 Cruz, California.
- 22 McBain and Trush (eds.). 2002. *San Joaquin River Restoration Study Background*
23 *Report*. Prepared for Friant Water Users Authority, Lindsay, CA and the Natural
24 Resources Defense Council, San Francisco, CA.
- 25 McBain and Trush. 2004. *Coarse Sediment Management Plan for the Lower Tuolumne*
26 *River*. Revised Final Report. Pg 77.
- 27 McCabe, G. T., Jr., and C. A. Tracy. 1994. *Spawning and early life history of white*
28 *sturgeon, Acipenser transmontanus, in the lower Columbia River*. *Fisheries*
29 *Bulletin* 92:760–772.
- 30 McEwan, D.R. 2001. *Central Valley steelhead. Pages 1-43 in R. L. Brown, editor.*
31 *Contributions to the biology of Central Valley salmonids*. Fish Bulletin 179:
32 Volume 1. California Department of Fish and Game, Sacramento.
- 33 McEwan, D., and T.A. Jackson. 1996. *Steelhead restoration and management plan for*
34 *California. Management report*. California Department of Fish and Game, Inland
35 Fisheries Division, Sacramento, California.
- 36 Meehan, W. R., and T. C. Bjornn. 1991. *Salmonid distributions and life histories. In*
37 *influences of forest and rangeland management on salmonid fishes and their*
38 *habitats*. Edited by W. R. Meehan. American Fisheries Society Special
39 Publication No. 19. Bethesda, Maryland. Pages 47–82.

- 1 Meeuwig, M.H., J.M. Bayer, and J.G. Seelye. 2005. *Effects of temperature on survival*
2 *and development of early life stage Pacific and western brook lampreys.*
3 *Transactions of the American Fisheries Society* 134:19–27.
- 4 Meng, L., and P.B. Moyle. 1995. *Status of splittail in the Sacramento-San Joaquin*
5 *Estuary.* *Transactions of the American Fisheries Society* 124: 538–549.
- 6 Miller, K.M., A. Teffer, S. Tucker, S. Li, A.D. Schulze, M. Trudel, F. Juanes, A. Tabata,
7 K. Kaukinen, N.G. Ginther, T.J. Ming, S.J. Cooke, J.M. Hipfner, D.A. Patterson,
8 and S.G. Hinch. 2014. *Infectious disease, shifting climates, and opportunistic*
9 *predators: cumulative factors potentially impacting wild salmon declines.*
10 *Evolutionary Applications* 7: 812-855.
- 11 Mills, T.J., D. McEwan, and M.R. Jennings. 1997. *California salmon and steelhead:*
12 *beyond the crossroads.* Pages 91-111 in D. J. Strouder, P.A. Bison, and R.J.
13 Naiman, eds. *Pacific salmon and their ecosystems.* New York: Chapman and Hall.
- 14 Moore, J.W., and J.M. Mallatt. 1980. *Feeding of larval lamprey.* *Canadian Journal of*
15 *Fisheries and Aquatic Sciences* 37:1658–1664.
- 16 Moyle, P.B. 2002. *Inland fishes of California: revised and expanded.* University of
17 California Press, Berkeley.
- 18 Moyle, P.B. and R.D. Nichols. 1973. *Ecology of some native and introduced fishes of the*
19 *Sierra Nevada foothills in Central California.* *Copeia* 1973: 978–990.
- 20 Moyle, P. B., and D. M. Baltz. 1985. *Microhabitat use by an assemblage of California*
21 *stream fishes: developing criteria for instream flow determinations.* *Transactions*
22 *of the American Fisheries Society* 114:695-704.
- 23 Moyle, P.B., R.D. Baxter, T. Sommer, T.C. Foin, and S.A. Matern. 2004. *Biology and*
24 *population dynamics of Sacramento Splittail (Pogonichthys macrolepidotus) in*
25 *the San Francisco Estuary: a review.* *San Francisco Estuary and Watershed*
26 *Science* [online serial] 2(2):1-47.
- 27 Moyle, P. B., J. A. Isreal, and S. E. Purdy. 2008. *Salmon, steelhead, and trout in*
28 *California: status of an emblematic fauna.* Report commissioned by California
29 Trout Center for Watershed Sciences, University of California, Davis.
- 30 Moyle, P.B., R.M. Quinones, J.V. Katz, and J. Weaver. 2015. *Fish Species of Special*
31 *Concern in California, Third Edition.* California Department of Fish and Wildlife,
32 Sacramento, CA.
- 33 Mussen, T.D., D. Cocherell, Z. Hockett, A. Ercan, H. Bandeh, M.L. Kavvas, J.J. Cech,
34 and N.A. Fangue. 2013. *Assessing juvenile Chinook salmon behavior and*
35 *entrainment risk near unscreened water diversions: Large flume simulations.*
36 *Transactions of the American Fisheries Society* 142: 130-142.

- 1 National Oceanic and Atmospheric Administration National Marine Fisheries Service
2 (NMFS). 1996. *Factors for decline: a supplement to the notice of determination*
3 *for west coast steelhead under the Endangered Species Act*. National Marine
4 Fisheries Service, Protected Resource Division, Portland, Oregon, and Long
5 Beach California.
- 6 _____. 1999. *West Coast Chinook salmon fact sheet*. Protected Resources Division.
7 Portland, Oregon
- 8 _____. 2009. *Final biological opinion and conference opinion on the long-term central*
9 *valley project and State Water Project operations criteria and plan*. Endangered
10 Species Act Section 7 Consultation. Final Biological Opinion. June 4.
- 11 _____. 2014. *Recovery plan for the evolutionarily significant units of Sacramento River*
12 *winter-run Chinook salmon and Central Valley spring-run Chinook salmon and*
13 *the distinct population segment of California Central Valley steelhead*. California
14 Central Valley Area Office.
- 15 Nicholas, J.W., and D.G. Hankin. 1989. *Chinook salmon populations in Oregon coastal*
16 *river basins: descriptions of life histories and assessment of recent trends in run*
17 *strengths*, Report EM 8402-Oregon Department of Fish and Wildlife, Research
18 and Development Section, Corvallis, Oregon.
- 19 Nichols, K. 2002. Merced River PKD survey – Spring 2002. *Memorandum to the San*
20 *Joaquin River Basin fish health information distribution list*. U.S. Fish and
21 Wildlife Service, CA-NV Fish Health Center, Anderson, California. December 6.
- 22 Nichols, K., and J.S. Foott. 2002. *Health monitoring of hatchery and natural fall-run*
23 *Chinook salmon juveniles in the San Joaquin River and tributaries, April – June*
24 *2001*. FY 2001 Investigation Report by the U.S. Fish and Wildlife Service,
25 California-Nevada Fish Health Center, Anderson, California.
- 26 Osterback, A-M.K., D.M. Frechette, A.O. Shelton, S.A. Hayes, M.H. Bond, S.A. Shaffer,
27 and J.W. Moore. 2013. *Hig predation on small populations: avian predation on*
28 *imperiled salmonids*. *Ecosphere* 4(9):116.
- 29 Pagliughi, S.P. 2008. *Lower Mokelumne River Reach specific thermal tolerance criteria*
30 *by life stage for fall-run Chinook salmon and winter-run steelhead*. East Bay
31 Municipal Utility District. Unpublished Report. 91pp.
- 32 Panshin, S.Y., N.M. Dubrovsky, J.M. Gronberg, and J.L. Domagalski. 1998. *Occurrence*
33 *and distribution of dissolved pesticides in the San Joaquin River Basin,*
34 *California*. Water-Resources Investigations Report 98-4032. U.S. Geological
35 Survey, National Water-Quality Assessment Program, Sacramento, California.
- 36 Peven, C.M., R.R. Whitney, and K.R. Wouldiams. 1994. *Age and length of steelhead*
37 *smolts from the mid-Columbia River basin, Washington*. *North American Journal*
38 *of Fisheries Management* 14: 77–86.

- 1 Quinn, N.W.T., Karkoski, J., 1998. *RealPotentialPotential for real time management of*
2 *water quality in the San Joaquin Basin, California*. Journal of the American
3 Water Resources Association 34 (6).
- 4 Rasmussen, D., B.A. Agee, P.T. Phillips. 1995. *Toxic substances monitoring program*
5 *1985*. California State Water Resources Control Board, Water Quality Monitoring
6 Report No 87-1WQ, Sacramento, CA 51 pp + Appendices.
- 7 Reynolds, F. L., T. J. Mills, R. Benthin, and A. Low. 1993. *Restoring Central Valley*
8 *streams: a plan for action*. California Department of Fish and Game, Inland
9 Fisheries Division. Sacramento, California.
- 10 Roni, P., T. Beechie, G. Pess, and K. Hanson. 2015. *Wood placement in river restoration:*
11 *fact, fiction, and future direction*. Canadian Journal of Fisheries and Aquatic
12 Sciences 72: 466-478.
- 13 Rutter, C. 1908. *The fishes of the Sacramento-San Joaquin basin, with a study of their*
14 *distribution and variation*, Bulletin of the U. S. Bureau of Fisheries 27: 103–152.
- 15 Sabal, M., Hayes, S., Merz, J. and Setka, J., 2016. *Habitat Alterations and a Nonnative*
16 *Predator, the Striped Bass, Increase Native Chinook Salmon Mortality in the*
17 *Central Valley, California*. North American Journal of Fisheries Management,
18 36(2), pp.309-320.
- 19 Saiki, M.K. 1984. *Environmental conditions and fish faunas in low elevation rivers on*
20 *the irrigated San Joaquin Valley floor, California*. California Fish and Game 70:
21 145-157.
- 22 _____. 1989. *Selenium and other trace elements in fish from the San Joaquin Valley and*
23 *Suisun Bay, 1985*. In: Howard AQ (ed) Selenium and Agricultural Drainage:
24 Implications for San Francisco Bay and the California Environment (Selenium
25 IV). The Bay Institute of San Francisco, Tiburon, CA, pp 35-49.
- 26 Saiki, M.K., T.P. Lowe. 1987. *Selenium in aquatic organisms from subsurface*
27 *agricultural drainage water, San Joaquin Valley, California*. Archives of
28 Environmental Contamination and Toxicology 16: 657-670.
- 29 Saiki, M.K., T.W. May. 1988. *Trace element residues in bluegills and common carp from*
30 *the lower San Joaquin River, California and its tributaries*. Science of the Total
31 Environment 74: 199-217.
- 32 Saiki, M.K. and D.U. Palawski. 1990. *Selenium and other elements in juvenile striped*
33 *bass from the San Joaquin Valley and San Francisco Estuary, California*.
34 Archives of Environmental Contamination and Toxicology 19: 717-730.
- 35 Saiki, M.K., M.R. Jennings, R.H. Wiedmeyer. 1992. *Toxicity of agricultural subsurface*
36 *drainwater from the San-Joaquin Valley, California, to juvenile Chinook salmon*
37 *and striped bass*. Transactions of the America Fisheries Society 121: 78-93.

- 1 San Joaquin River Restoration Program (SJRRP). 2010a. *Fisheries management plan: a*
2 *framework for adaptive management in the San Joaquin River Restoration*
3 *Program*. San Joaquin River Restoration Program. November 2010.
- 4 _____. 2010b. *Conceptual models of stressors and limiting factors for San Joaquin River*
5 *Chinook salmon*. Exhibit A of fisheries management plan. November 2010.
- 6 _____. 2011a. *Programmatic biological assessment. San Joaquin River Restoration*
7 *Program*. November 2011.
- 8 _____. 2011b. *San Joaquin River Restoration Program. Appendix B. 2011 Draft Annual*
9 *Technical Report*. Fish passage evaluation. Task 1, evaluation of partial fish
10 passage barriers. July 2011.
- 11 _____. 2011c. *2010 Annual Technical Report: Appendix D Surface Water Quality*. April
12 2011.
- 13 _____. 2012. *San Joaquin River Restoration Program Fish Passage Evaluation. Task 2,*
14 *Draft Technical Memorandum, Evaluation of Partial Fish Passage Barriers*. July
15 2012.
- 16 _____. 2013. *Chinook salmon trap and haul summary*.
- 17 _____. 2014. *2013-2014 Fish Assemblage Inventory and Monitoring*. 2014 Mid-Year
18 Technical Report. Prepared for SJRRP by USFWS, Stockton, CA.
- 19 _____. 2016. *Hatchery and Genetic Management Plan*. Prepared for the SJRRP by the
20 UC Davis Genomic Variation Lab, Davis, California and CDFW, Fresno,
21 California.
- 22 _____. 2017. *Fish Assemblage Inventory and Monitoring 2013-2014. Draft Final*
23 *Monitoring and Analysis Plan Report*. Prepared for San Joaquin River Restoration
24 Program by U.S. Bureau of Reclamation, Technical Service Center, Denver,
25 Colorado and USFWS, Lodi Fish and Wildlife Office, Lodi, California.
- 26 Sellheim, K. L., C. B. Watry, B. Rook, S. C. Zeug, J. Hannon, J. Zimmerman, K. Dove,
27 and J. E. Merz. 2015. *Juvenile Salmonid Utilization of Floodplain Rearing Habitat*
28 *after Gravel Augmentation in a Regulated River*. River Research and Applications
29 32(4):610-621.
- 30 Shapovalov, L., and A.C. Taft. 1954. *The life histories of the steelhead rainbow trout*
31 *(Salmo gairdneri gairdneri) and silver salmon (Oncorhynchus kisutch) with*
32 *special reference to Waddell Creek, California, and recommendations regarding*
33 *their management*. California Department of Fish and Game Fish Bulletin 98.
- 34 Sogard, S.M., Merz, J.E., Satterthwaite, W.H., Beakes, M.P., Swank, D.R., Collins, E.M.,
35 Titus, R.G. and Mangel, M. 2012. *Contrasts in habitat characteristics and*
36 *life history patterns of Oncorhynchus mykiss in California's central coast*

- 1 *and Central Valley*. Transactions of the American Fisheries Society, 141(3),
2 pp.747-760.
- 3 Sommer, T., R. Baxter, and B. Herbold. 1997. *Resilience of splittail in the Sacramento-*
4 *San Joaquin estuary*. Transactions of the American Fisheries Society 126: 961–
5 976.
- 6 Sommer, T.R., Harrell, W.C., and Nobriga, M.L. 2005. *Habitat use and stranding risk of*
7 *juvenile Chinook salmon on a seasonal floodplain*. North American Journal of
8 Fisheries Management 25: 1493-1504.
- 9 Sommer, T.R., M.L. Nobriga, W.C. Harrell, W. Batham, and W.J. Kimmerer. 2001.
10 *Floodplain rearing of juvenile chinook salmon: evidence of enhanced growth and*
11 *survival*. Canadian Journal of Fisheries and Aquatic Sciences 58: 325-333.
- 12 S.P. Cramer and Associates. 2004. 2002-04 *Stanislaus River weir data report*. Final
13 report prepared for the Anadromous Fish Restoration Program. October.
- 14 _____. 2005. 2004-05 *Stanislaus River weir data report*. Final report prepared for the
15 Anadromous Fish Restoration Program. June.
- 16 Steinbach Elwell, L.C., K.E. Stromberg, E.K.N. Ryce, and J. Bartholomew. 2009.
17 *Whirling Disease in the United States: A Summary of Progress in Research and*
18 *Management*. Prepared for the Whirling Disease Initiative of the Montana Water
19 Center at Montana State University, Bozeman, Montana. 61 p.
- 20 Strange, J.S. 2010. *Upper thermal limits to migration in adult Chinook Salmon: Evidence*
21 *from the Klamath River basin*. Transactions of the American Fisheries Society
22 139: 1091-1108.
- 23 Tatara, C. P. and B.A. Berejikian. 2012 *Mechanisms influencing competition between*
24 *hatchery and wild juvenile anadromous Pacific salmonids in fresh water and their*
25 *relative competitive abilities*. Environmental Biology of Fishes 94:7-19.
- 26 The Bay Institute of San Francisco. 1998. *From the Sierra to the sea: The ecological*
27 *history of the San Francisco Bay-Delta watershed*. Copyright 1998 The Bay
28 Institute of San Francisco.
- 29 Tucker, M.E., Wouldiams, C.M. and Johnson, R.R. 1998. *Abundance, food habits and*
30 *life history aspects of Sacramento squawfish and striped bass at the Red Bluff*
31 *Diversion Complex, including the research pumping plant, Sacramento River,*
32 *California, 1994-1996*. Red Bluff Research Pumping Plant Report Series, Volume
33 4. U.S. Fish and Wildlife Service, Red Bluff, California.
- 34 United States Environmental Protection Agency (EPA). 2003. *EPA Region 10 guidance*
35 *for Pacific Northwest state and tribal temperature water quality standards*. EPA
36 910-B-03-002. Region 10 Office of Water, Seattle, WA.

- 1 United States Fish and Wildlife Service (USFWS). 1992. Shaded riverine aquatic cover
2 of the Sacramento River System: Classification as resource category 1 under the
3 FWS mitigation policy. USFWS Sacramento Field Office, Sacramento, CA.
- 4 _____. 1998. Identification of the instream flow requirements for anadromous fish in the
5 streams within the central valley of California. Annual progress report fiscal year
6 1998. USFWS Sacramento Fish and Wildlife Office, Sacramento, CA.
- 7 _____. 2010. Flow-habitat relationships for juvenile fall/spring-run Chinook salmon and
8 steelhead/rainbow trout rearing in the Yuba River. USFWS Sacramento Fish and
9 Wildlife Office, Energy Planning and Instream Flow Branch, Sacramento, CA.
- 10 _____. 2011. Flow-habitat relationships for juvenile spring-run Chinook salmon and
11 steelhead/rainbow trout rearing in Clear Creek between Whiskeytown Dam and
12 Clear Creek Road. USFWS Sacramento Fish and Wildlife Office, Restoration and
13 Monitoring Program, Sacramento, CA.
- 14 Van de Wetering, S.J. 1998. *Aspects of life history characteristics and physiological*
15 *processes in smolting pacific lamprey (Lampetra tridentata) in a central Oregon*
16 *coast stream. Master of Science Thesis. Oregon State University. Corvallis,*
17 *Oregon 59 pp.*
- 18 Vernier, J.M. 1969. *Chronological table of embryonic development of rainbow trout,*
19 *Canada Fisheries and Marine Service Translation Series 3913.*
- 20 Viant, M.R., C.A. Pincetich, and R.S. Tjeerdema. 2006. *Metabolic effects of dinoseb,*
21 *diazinon and esfenvalerate in eyed eggs and alevins of Chinook salmon*
22 *(Oncorhynchus tshawytscha) determined by H1 NMR metabolomics. Aquatic*
23 *Toxicology 77:359-371.*
- 24 Voight, H.N., and D.B. Gale. 1998. *Distribution of fish species in tributaries of the lower*
25 *Klamath River: an interim report, FY 1996.* Technical report, No. 3. Yurok Tribal
26 Fisheries Program, Habitat Assessment and Biological Monitoring Division.
- 27 Wagner, H. H., R. L. Wallace, and H. K. Campbell. 1963. *The seaward migration and*
28 *return of hatchery-reared steelhead trout in the Alsea River, Oregon.*
29 *Transactions of the American Fisheries Society 92:202–210.*
- 30 Wang, J.C.S. 1986. *Fishes of the Sacramento-San Joaquin estuary and adjacent waters,*
31 *California: a guide to the early life histories.* Technical Report 9. Prepared for the
32 Interagency Ecological Study Program for the Sacramento-San Joaquin Estuary
33 by California Department of Water Resources, California Department of Fish and
34 Game, U. S. Bureau of Reclamation and U. S. Fish and Wildlife Service.
- 35 _____. 1995. *Observations of early life stages of splittail (Pogonichthys*
36 *macrolepidotus) in the Sacramento-San Joaquin estuary, 1988 to 1994.* IEP
37 Technical Report 43.

- 1 Wang, S., J.J. Hard, and F. Utter. 2002. *Salmonid inbreeding: a review. Reviews in Fish*
2 *Biology and Fisheries 11:301-319.*
- 3 Ward, B. R., and P. A. Slaney. 1988. *Life history and smolt-to-adult survival of Keogh*
4 *River steelhead trout (Salmo gairdneri) and the relationship to smolt size.*
5 *Canadian Journal of Fisheries and Aquatic Sciences 45:1110–1122.*
- 6 Ward, P.D., T.R. McReynolds, and C.E. Garman. 2003. *Butte and Big Chico creeks*
7 *spring-run Chinook salmon, Oncorhynchus tshawytscha, life history investigation*
8 *2001-2002.* Inland Fisheries Administrative Report, California Department of
9 Fish and Game, Chico, CA.
- 10 Weber, E.D. and K.D. Fausch. 2005. *Competition between hatchery-reared and wild*
11 *juvenile Chinook Salmon in enclosures in the Sacramento River, California.*
12 *Transactions of the American Fisheries Society 134: 44-58.*
- 13 White, J. R., P. S. Hoffmann, K., and S. Baumgartner. 1988. *Selenium verification study,*
14 *1986–1987.* A report to California State Water Resources Control Board from
15 California Department of Fish and Game, Sacramento, CA, 60 pp + Appendices.
- 16 Wiese, F. K., Parrish, J. K., Thompson, C. W., and Maranto, C. 2008. *Ecosystem-based*
17 *management of predator-prey relationships: piscivorous birds and salmonids.*
18 *Ecological Applications 18(3): 681-700.*
- 19 Yoshiyama, R.M., E.R. Gerstung, F.W. Fisher, and P.B. Moyle. 1996. *Historical and*
20 *present distribution of chinook salmon in the Central Valley drainage of*
21 *California, Sierra Nevada Ecosystem Project: final report to congress, Volume*
22 *III: Assessments, commissioned reports, and background information.* University
23 of California, Center for Water and Wildland Resources, Davis, California, pp.
24 309-362.
- 25 Yoshiyama, R.M., F.W. Fisher, and P.B. Moyle. 1998. *Historical abundance and decline*
26 *of Chinook salmon in the Central Valley region of California.* *North American*
27 *Journal of Fisheries Management 18: 487-521.*
- 28 Young, P.S., and J.J. Cech, Jr. 1996. *Environmental tolerances and requirements of*
29 *splittail.* *Transactions of the American Fisheries Society 125: 664–678.*